



Analysis of specific pathogens as possible triggers of Multiple Sclerosis

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I. Overview

Multiple sclerosis (MS) is believed, based on familial concordance data, to be a multifactorial disorder, requiring the presence of both genetic and environmental factors to initiate the mechanisms that lead to demyelination and neural damage. Although the environmental factors that trigger MS are still being determined, infectious agents such as bacteria and viruses have long been speculated to play a role in the etiology of MS. Scientists have investigated many different pathogens as candidate triggers of MS, using a variety of experimental techniques that have evolved over time as knowledge about infectious agents and disease has grown.

This document surveys a number of pathogens that have been evaluated for a role in MS, summarizing the results of the experiments that have been conducted to identify possible associations. It does not provide an exhaustive analysis for each pathogen; detailed reviews already exist for a number of them. Instead, it attempts to give an overview of the types of investigations that have been conducted across the spectrum of infectious agents explored in MS, and provide a sense of what is known and not known regarding the involvement of these potential infectious triggers of MS.

Each pathogen discussed in this document is presented in a standardized template. The templates contain a brief description of the pathogen being evaluated and a summary of the available experimental evidence, primarily obtained from case-control or epidemiological studies. The evidence is organized into these topics:

- Is the presence of the actual pathogen different in subjects with MS compared with non-MS subjects?
- Does the immune response to the pathogen differ between MS subjects and non-MS subjects?
- Does the age of infection or another characteristic of infection differ significantly between MS subjects and non-MS subjects?
- Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?
- If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?

At the end of each template, an opinion is given as to the strength of the evidence associating the pathogen with the risk of MS.

For most of the pathogens listed, few conclusions can be drawn about their possible involvement in MS. A common finding is that positive results provided by one or more studies are contradicted by another study or group of studies.

There are several possible reasons for this situation, including:

- Differences and/or deficiencies in the experimental methods and techniques used (see Moore and Wolfson¹ for an analysis of how poor methodological choices or incomplete descriptions of methods limit reliance on results)
- Differences in choices of cases and control subjects and differences in recruitment methods used
- Small sample sizes resulting in a lack of statistical power
- Inherent difficulties in detecting or isolating certain pathogens (such as *Chlamydia pneumoniae*), which may prevent replication of results from lab to lab
- Non-specific findings implicating multiple pathogens (none, one, or some of which may be true susceptibility factors)
- Lengthy time spans between the age of initial infection, the age of MS onset, and the age at which tissues or data are collected for analysis; the infection-to-analysis span can be especially long for studies examining MS autopsy brain tissue
- The possibility that MS may be a etiologically heterogeneous disease and therefore evidence for any given infectious trigger may be subtle or modest, or not even present in a given population under study
- The possibility that differences detected between MS and non-MS subjects may be artifacts of the disease process (for instance, the presence of pathogens in the central nervous system in MS subjects may be an artifact of blood-brain barrier leakage)

Indeed, because of time lapses, disease heterogeneity, and other factors, it is probably difficult to conclusively *exclude* the involvement of any particular pathogen of interest in any type or form of MS. Still, we believe there is value in presenting the available data as it will help to highlight areas where additional research could more conclusively indicate whether infections trigger MS and if so, which pathogens are involved.

Note: Accompanying this document is a spreadsheet (continually being expanded) that lists relevant studies for each pathogen, including characteristics such as technique used, number of subjects and controls, and results. A copy of this spreadsheet can be downloaded at www.acceleratedcure.org/downloads/phase2-infectiousagents-studies.xls.

II. Viruses

Pathogen: Adenovirus
<i>Description:</i>
Adenoviruses are common human pathogens that can cause respiratory infections, conjunctivitis, and occasionally gastroenteritis. These viruses most frequently infect children although outbreaks in adult groups are also possible. Adenoviruses persist in human tonsils and are capable of integrating their genome into the host DNA, making them candidate vectors for gene therapy.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
Evidence is limited – one study ² found adenovirus-like particles in brain tissue of 1 of 20 MS subjects but these were not correlated with demyelinating lesions; another study ³ found no adenovirus DNA in MS brain tissue.
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
The few studies found that compared adenovirus antibodies in MS subjects and controls (e.g., Appel <i>et al</i> ⁴) did not find a significant difference between the two groups.
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
Not assessed
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
Not assessed
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
Evidence is limited – one study ⁵ found a correlation between upper respiratory infections (URIs) associated with increased cerebrospinal fluid (CSF) adenovirus titers and subsequent MS relapses.
<i>Conclusion:</i>
Only a few studies were found that explore a causal role for this virus in MS. The correlation found between adenovirus titers, URIs and subsequent relapses could indicate a possible interaction between the presence and activity of adenoviruses and MS disease mechanisms.

Pathogen: Canine distemper virus (CDV)
<i>Description:</i>
CDV is a virus that infects dogs and other animals, including wildlife, and can result in encephalomyelitis. It is closely related to the measles virus. Humans can be infected with CDV asymptotically, but the measles vaccine protects humans against CDV infection. A vaccine against CDV was introduced in the 1960's and is now widely available for pet dogs.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
Evidence so far is unresponsive. One tissue staining study showed features characteristic of CDV ⁶ in MS plaques; however, subsequent studies ⁷⁻⁹ showed no evidence of CDV-specific genes or antigens in MS brain samples.
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
Evidence is inconclusive: <ul style="list-style-type: none"> • A few studies found an increase in anti-CDV antibodies in MS serum and/or CSF compared with controls, but increases in measles antibodies were also seen (e.g., Cook <i>et al</i>¹⁰). Other similar studies found no significant difference in antibody levels between MS and control subjects (e.g., Kurtzke <i>et al</i>¹¹), and another study showed no IgM response to CDV in MS CSF.¹² • No difference was seen between MS subjects and controls in lymphocyte production upon exposure to CDV.¹³
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
Some epidemiological studies have shown connections between CDV outbreaks, dog ownership, etc. and MS; however, others have found no such association. A review of CDV epidemiological studies ¹⁴ concludes that weaknesses in these studies have prevented demonstration of a relationship, if any exists.
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
As mentioned, immunization for dogs is now routine in many countries which has significantly reduced the incidence of CDV. However, no corresponding reduction in the risk of MS has been demonstrated.
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
CDV is known to cause demyelination in animals, and one study showed a loss of beta-adrenergic receptors on astrocytes in subjects with MS as well as CDV-infected dogs which may indicate involvement of a common pathogenic factor. ¹⁵
<i>Conclusion:</i>
There is no strong evidence at this time to support a role for CDV as a cause of MS. If CDV were an important trigger of MS, it would be expected that control of this virus would produce an eventual reduction in the incidence of MS, but such a reduction has not yet been documented.

Pathogen: Coronavirus
<i>Description:</i>
Human coronaviruses (HCV) are responsible for approximately 20 percent of colds in humans. Incidence of coronavirus infection peaks in the winter. Immunity does not persist and individuals can be periodically reinfected. There are two main serotypes of HCV, OC43 and 229E. While human coronaviruses typically infect respiratory epithelial cells, they have been found in brain tissue as well, and the coronavirus murine hepatitis virus (MHV) is known to cause a chronic demyelinating disease similar to MS in mice.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
Studies of the presence of coronavirus in brain tissue have provided mixed results. Initial reports of coronavirus or particles resembling coronavirus in some MS brains ^{16, 17} motivated several subsequent efforts to analyze the presence of coronavirus in brain tissue. Some (e.g., Stewart <i>et al</i> ¹⁸ and Arbour <i>et al</i> ¹⁹) detected coronavirus RNA (either OC43 or 229E) more frequently in brain tissue of MS subjects compared with controls, but others such as Dessau <i>et al</i> ²⁰ found no statistical difference between the two groups.
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
Evidence so far is unresponsive. Several studies have reported finding no difference in serum antibody titers between MS and controls. Results are mixed from the few studies that examined whether CSF titers to coronavirus are elevated in MS. ²¹⁻²³
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
Not assessed
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
Not assessed
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
One study found that MS subjects had more HCV 229E/MBP cross-reactive T cells than did controls; ²⁴ this may suggest molecular mimicry which could contribute to the development of MS, or may instead be a result of greater access to myelin antigens in MS. Another study demonstrated that HCV infection of human astrocytes can spur an increase in the production of inflammatory molecules which may be damaging to brain tissue. ²⁵ Also, as mentioned above, the mouse coronavirus MHV does cause a form of demyelination similar to MS.
<i>Conclusion:</i>
At this time no strong evidence has emerged to support a causal role for coronavirus in MS. Once the role of MHV in demyelination is better understood, it may be possible to compare it with HCV to determine which of MHV's pathogenic factors are also present in HCV.

Pathogen: Cytomegalovirus
<i>Description:</i>
Human cytomegalovirus (CMV or HCMV) is a member of the herpes family and is a common pathogen worldwide. It causes three types of clinical syndromes: (1) congenital infection can cause hepatosplenomegaly, retinitis, rash, and CNS involvement, although 90% of infected infants are asymptomatic; (2) primary infections in older children and adults can cause mononucleosis, although 90% of primary infections at this age are asymptomatic, and (3) primary or reactivated infections in immunocompromised individuals (e.g., people with AIDS) can develop into a life-threatening systemic infection involving the CNS, lungs, GI tract, liver and retinas. Transmission occurs via contact with infected secretions such as saliva or breast milk. HCMV can persist in and be chronically excreted by a variety of tissues, including salivary glands and blood cells.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
Evidence so far is unresponsive: <ul style="list-style-type: none"> • Three studies found no HCMV DNA or antigen in MS brain tissue,^{3, 26, 27} while another found similar percentages of MS and control subjects (neurological and non-neurological) positive for HCMV DNA in brain tissue.²⁸ • Of those studies searching for HCMV DNA in other tissues, one found it present in the PBMCs of MS subjects but not healthy controls.²⁹ However, another found no difference in HCMV DNA prevalence in PBMCs of MS subjects and controls,³⁰ while others failed to detect any HCMV DNA in CSF or serum samples from MS subjects or controls.³⁰⁻³²
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
Again, the available evidence is unresponsive: <ul style="list-style-type: none"> • Most studies of anti-HCMV antibodies using serum or CSF have found similar levels or prevalence in MS and control subjects (e.g., Alvarez-Lafuente <i>et al</i>,³⁰ Ascherio <i>et al</i>,³³ and Alotaibi <i>et al</i>³⁴). • Similarly, other types of immunological studies have found either no HCMV-specific differences or similar findings in MS and control subjects (e.g., Haahr <i>et al</i>³⁵, Hollsberg <i>et al</i>³⁶ and Lunemann <i>et al</i>³⁷). • Scotet <i>et al</i>³⁸ detected CMV- and/or EBV-reactive T cells in lesions from a variety of different inflammatory and/or autoimmune diseases, suggesting an involvement of antiherpes immune factors in inflammation that may be not specific to MS. • Interestingly, one study (Zivadinov, <i>et al</i>³⁹) reported that more favorable clinical and MRI outcomes in MS subjects were associated with HCMV antibody positivity and higher titers.
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
Not assessed
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
Not assessed
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
One study ⁴⁰ that investigated viral activity during acute MS attacks reported that HCMV DNA was found in the blood of some MS subjects on the first day of the acute attack

and during follow-up. However, other viruses were detected as well.

Conclusion:

Although its persistence and potential for CNS involvement make it a plausible candidate for an MS trigger, at this time little evidence exists to support a specific role for HCMV in causing MS.

Pathogen: Epstein-Barr virus**Description:**

Epstein-Barr virus (EBV) is a ubiquitous member of the herpes family; 95% of the world's population is thought to have been infected with this virus. Primary EBV infection can be asymptomatic or can manifest as infectious mononucleosis (IM), especially in adolescents and young adults. (Primary infection in children is often mild or asymptomatic.) Development of IM during primary infection is associated with large expansions of T cells in the blood. Why IM develops in some people but not others is not known but it may be due to host factors (such as genetics) and/or characteristics of the infection. EBV is trophic for B cells and is thought to cause lymphoproliferation in immunocompromised individuals. EBV persists in B cells and epithelial cells as a latent infection that periodically reactivates, and is primarily exchanged through transmission of saliva.

Is presence of actual pathogen different in MS subjects compared with non-MS subjects?

Several studies have looked into this question and provide a range of findings:

- **Brain tissue:** Six studies have been conducted to investigate the presence of EBV DNA in brain tissue. Of these, Hilton *et al*⁴¹ and Virtanen *et al*⁴⁷ found no evidence of EBV RNA in MS plaques, Sanders *et al*⁴⁸ reported that EBV DNA was not more prevalent in MS samples than controls, and Challoner *et al*⁴⁹ and Morre *et al*⁵⁰ detected no EBV DNA in MS brain. However, a recent study by Serafini *et al*⁵¹ found EBV-infected B cells in 21 of 22 MS brain samples studied (no infection was found in samples from subjects with other neurological diseases). EBV-infected cells were found both in active lesions and in ectopic B cell follicles located in cerebral meningeal tissue.
- **CSF:** Neither Martin *et al*⁵² nor Morre *et al*⁵⁰ detected EBV in the CSF of any MS subjects. Serafini *et al*⁵¹ detected low copy numbers of EBV DNA in the CSF of only 2 of 16 MS cases, Mancuso *et al*⁵³ found similar prevalence of EBV DNA in MS subjects and controls; and Alvarez-Lafuente *et al*⁵⁴ found EBV DNA in the CSF of only 1 of 48 MS subjects.
- **Blood/serum:** Several analyses of EBV DNA have been conducted using the blood and/or serum of MS subjects and controls, with mixed results:
 - EBV was found in none of the subjects or controls in Martin *et al*⁵², Villoslada *et al*⁵⁵, or Riverol *et al*⁵⁶ (serum).
 - EBV was found in all subjects and controls in Hay and Tenser⁴⁷ (PBLs)
 - EBV was found in equivalent percentages of subjects and controls by Ferrante *et al*⁵⁷ (included both acute and stable MS), Alvarez-Lafuente *et al*⁵⁴, Lunemann *et al*⁵⁸, and Sotelo *et al*⁵⁹ (PBMCs); and Alvarez-Lafuente *et al*⁴⁹ (serum and PBMCs).
 - A prospective study⁵⁰, using samples drawn before onset or diagnosis, found that the presence of EBV in plasma (but not the viral load) was associated with an increased risk of MS.
 - The presence of EBV DNA was associated with MS exacerbations by Ferrante *et al*⁵⁷ (which detected EBV DNA in the PBMCs of 42.8% of MS subjects on day 1 of attack) and Wandinger *et al*⁶⁰ (which detected DNA in the serum of 72% of MS subjects with exacerbations vs. 0% of stable subjects). However, another study failed to find increased anti-EA-D IgG (a sign of viral reactivation) in MS subjects during relapse as compared with baseline.⁵²

- Level of disease activity has also been associated with the presence of EBV. Fraser *et al*⁶³ established spontaneous B lymphocyte transformation from 8 of 10 active MS subjects but only 3 of 18 stable MS subjects and only 4 of 20 healthy controls; all transformed cells carried EBV antigens. Hollsberg *et al*⁶⁴ also found that the presence of EBV DNA in MS samples was associated with disease activity.

Does immune response to pathogen differ between MS subjects and non-MS subjects?

A variety of studies concerning the immunological response to EBV have been conducted with respect to MS; most have found some type of association with MS although not all of these results have been consistently verified:

- Of those studies examining antibody titers or seropositivity to EBV, a preponderance found either higher titers or a higher prevalence of seropositivity in MS or optic neuritis subjects vs. controls (although in some studies, these effects were found for other pathogens as well). Notably, Alotaibi *et al*⁶⁴ and Banwell *et al*⁶⁵ showed significantly higher seropositivity in MS pediatric subjects than in matched controls (83% vs. 42% and 86% vs. 64%, respectively).
- In addition, five studies^{33, 56-59} associated higher titers against EBV with the subsequent development of MS. The presence of higher antibody titers to EBNA-1 and EBNA complex prior to the development of MS may indicate a severe or recent primary infection or reinfection that stimulates a vigorous cellular immune response⁵⁸.
 - Note however that Sundstrom *et al*⁶⁷ suggests that higher pre-onset titers may be due to the development of proteasomal autoantibodies that can occur during MS, which may block EBV's ability to hide from immune system and make it more immunogenic.
- Haahr *et al*⁶⁰ found equal antibody evidence of EBV reactivation in MS subjects vs. controls, but reactivation was found more frequently in recently diagnosed MS subjects than those of longer disease duration.
- Two separate studies^{61, 62} investigated the interaction between the HLA DR15 gene and anti-EBNA1 titers in influencing MS risk; each found significantly increased MS risk in DR15-positive individuals with high antibody titers.
- Rand *et al*⁶³ found an oligoclonal banding pattern specific for the EBV antigen EBNA-1 in 5 of 15 MS subjects but 0 of 12 controls; Cepok *et al*⁶⁴ also found that oligoclonal bands from MS subjects specifically bound EBNA-1 and BRRF2, another EBV protein. Serafini *et al*⁶³ found EBV-specific OCBs in seven of 16 MS subjects studied; when present, OCBs were generally few in number and faint.
- Three studies⁶⁵⁻⁶⁷ identified a failure of T cells from MS subjects to control EBV infection of cells, which may be related to an increased CD4+/CD8+ ratio as opposed to overall numbers of CD8+ cells or T-cell mediated cytotoxicity.
- On the other hand, Lunemann *et al*⁶⁷ determined that EBNA1-reactive CD4+ cells from MS subjects were more numerous, produced more IFN-gamma, and recognized a broader range of EBNA1 antigens than cells from controls. A follow-up study demonstrated that EBNA-1-reactive CD4+ cells from MS subjects were more likely to also react to myelin antigens than to other autoantigens not associated with MS.⁶⁸
- Hollsberg *et al*⁶⁶ found a higher frequency of CD8+ cells responding to two EBV antigens, EBNA-3A and latent membrane protein 2 (but not to other EBV antigens and not to two CMV epitopes) in MS subjects vs. healthy controls.

<p>Cepok <i>et al</i>⁶⁴ found greater CD8+ T cell responses to latent EBV proteins in MS subjects than in non-MS neurological controls. Similarly, Jilek <i>et al</i>⁶⁹ described an increased frequency of IFN-gamma secreting EBV-specific CD8+ cells in subjects with a clinically isolated syndrome; this T-cell response tended to decline over the course of a year. However, no differences in the prevalence or levels of EBV-reactive cytotoxic lymphocytes were found between MS cases and controls in a study by Gronen, <i>et al</i>⁷⁰. In addition, as mentioned in the HCMV template above, Scotet <i>et al</i>⁸⁸ detected CMV and/or EBV-reactive CD8+ T cells in lesions from a variety of different inflammatory and/or autoimmune diseases, suggesting an involvement of antiherpes immune factors in inflammation that may be not specific to MS.</p>
<p><i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i></p>
<p>Numerous sources indicate that MS is associated with a history of infectious mononucleosis,^{60, 71-80} and that primary (initial) infection is absent in MS subjects, presumably because of previous exposure.^{51, 81, 82} One study, Munch <i>et al</i>⁸¹, investigated EBV subtypes, finding all 8 members of an MS cluster in Denmark to harbor the same EBV subtype. However, another study did not find any particular strain of EBV to be associated with MS.⁸³</p>
<p><i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i></p>
<p>The effect of anti-herpes treatment on MS disease activity was examined by Bech <i>et al</i>,⁸⁴ who found that valacyclovir appeared to reduce new active lesion formation in a subset of subjects who had active lesions at baseline. Another trial of valacyclovir found evidence of clinical efficacy in more severely affected subjects but detected no effect on imaging evaluation⁸⁵. Administration of valacyclovir in MS subjects appeared in one study to reduce EBV expression in saliva but not in plasma⁵⁴.</p>
<p><i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i></p>
<p>Several different types of evidence have been produced to explain how EBV could cause or trigger MS:</p> <ul style="list-style-type: none"> • Multiple reports of immune cross-reactivity to EBV and self-antigens have been published: <ul style="list-style-type: none"> ○ van Sechel <i>et al</i>⁸⁶ showed that EBV-infected B cells produce alpha beta crystallin which is an immunodominant antigen of CNS myelin in MS subjects, and Rand <i>et al</i>⁸⁷ found that oligoclonal bands (OCBs) in some MS subjects and neurological controls reacted to epitopes that are found both in EBNA and alpha beta crystallin. Unlike other stress proteins, alpha beta crystallin is not expressed in lymph nodes so it may evade tolerance mechanisms. ○ Esposito <i>et al</i>⁸⁸ demonstrated antibody cross-reactivity between human transaldolase epitope (271-285) and an EBV as well as an HSV-1 peptide. ○ Vaughan <i>et al</i>⁸⁹ detected in MS subjects cross-reactive antibodies to EBNA (Epstein-Barr nuclear antigen) and lymphocyte and neuroglial proteins. ○ Lang <i>et al</i>⁹⁰ found T-cell receptor (TCR) contact surface similarities between DRB5*0101-EBV (DNA polymerase peptide) and DRB1*1501-MBP(85-99). • Several studies have explored possible connections between EBV and

<p>retroviruses that may relate to MS. Haahr <i>et al</i>,⁹¹ Munch <i>et al</i>,⁹² Christensen <i>et al</i>,⁹³ and Christensen <i>et al</i>⁹⁴ established B cell lines from MS subjects that produced retrovirus-like particles as well as EBV; they also detected reverse transcriptase activity via PCR.</p> <ul style="list-style-type: none">• Pender⁹⁵ suggested that EBV-infection of autoreactive B cells lodging in the central nervous system may promote the survival of activated autoreactive T cells which then effect an inflammatory response leading to tissue damage. Subsequently, a study described EBV-infected B cells in the brain and meningeal tissue of MS subjects and also detected the presence of activated CD8+ T cells and evidence of cytotoxic activity in association with the infected cells.⁴³
<p><i>Conclusion:</i></p> <p>Numerous types of evidence exist to suggest a possible role for EBV in causing MS, including several findings that suggest possible etiological mechanisms. It is possible that some evidence (e.g., increased antibody titers in MS cases vs. controls) results from MS disease processes rather than reflects a cause of MS. However, the existence of multiple types of evidence supporting a causal role, particularly the increased seroprevalence of EBV in pediatric MS cases³⁴ and the detection of EBV-infected B cells in lesions and meningeal tissues,⁴³ is striking and makes EBV a pathogen of particular interest in MS. Questions worth exploring further include why mononucleosis appears to increase the risk of MS, what is the full significance of EBV-infected B cells in the central nervous system, and whether interactions between EBV and other elements such as endogenous retroviruses play a key role in MS.</p>

Pathogen: Herpes simplex virus
<i>Description:</i>
Herpes simplex virus (HSV) 1 and 2 are closely related neurotropic viruses whose primary symptoms are oral lesions (HSV-1) and genital lesions (HSV-2). Other manifestations of HSV infection include lesions at other skin sites, meningitis, encephalitis, and disseminated disease. HSV is transmitted by oral and sexual contact. Infection is established through replication of the virus in epithelial cells and movement up the peripheral sensory nerve to the dorsal ganglia, where it can further replicate, move back down the sensory nerves to form a new lesion, or assume a state of latency.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
<ul style="list-style-type: none"> • Several studies have searched for the presence of HSV in brain tissue, with varying results: <ul style="list-style-type: none"> ○ HSV antigen or DNA has been found in brain or plaque tissue of MS subjects and/or other subjects by Warren <i>et al</i>⁶⁶ (HSV was isolated from the trigeminal ganglia), Martin <i>et al</i>⁶⁷ (HSV-2 but not HSV-1 was found in 3 of 31 MS tissue samples, restricted to glial cell nuclei in and around demyelinating lesions), and Nicoll <i>et al</i>⁶⁸ (HSV-1 but not HSV-2 DNA was found in 1 of 77 plaques from 23 MS subjects). ○ Another study found no HSV DNA in MS brain tissue.²⁶ ○ In comparison with controls, results are slightly positive: see Challoner <i>et al</i>³ (HSV-1 was found in the brain tissue of 2 of 25 MS vs. 2 of 42 controls) and Sanders <i>et al</i>⁶⁹ (HSV was more often detected in MS brain tissue than OND or NND control tissue and was also more prevalent in active vs. inactive plaques). • Mixed results have also been found in studies of other tissues: <ul style="list-style-type: none"> ○ One study³¹ failed to detect HSV or other herpesvirus DNA in the serum or CSF of MS subjects; similarly, another study failed to detect herpes simplex virus DNA in the PBMCs of MS subjects (in relapse or remission) or controls⁴⁸. ○ Other studies examining PBMCs³⁰ and CSF^{32, 44} found HSV DNA in similar levels in MS subjects and controls. ○ HSV-1 or -2 DNA was found in the PBMCs of a subset of MS subjects at the beginning of relapses and during a follow-up period, but EBV and HTLV-1 were found more frequently.⁴⁰ ○ Similarly, HSV-1 mRNA and DNA were found in the PBMCs of acute MS subjects but not stable MS subjects or controls.²⁹
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
Evidence to date does not show significant differences between MS and non-MS subjects: <ul style="list-style-type: none"> • Antibody evidence so far is unresponsive. Most antibody studies of serum or CSF have reported similar titers or rates of positivity in MS and control subjects, although a few found higher titers or positivity rates in MS subjects, one using tear samples.¹⁰⁰ One study of pediatric MS subjects³⁴ showed lower positivity rates for MS subjects vs. controls. • In studies of T cell immunity to HSV, the majority showed no differences in lymphocyte response between MS and controls (e.g., Chou <i>et al</i>¹⁰¹), while a few showed lower responses to HSV by MS subjects (e.g., de Silva and

<p>McFarland¹⁰²). However, Brudek, <i>et al.</i> demonstrated increased IFN-gamma production in PBMCs from MS subjects vs. controls after stimulation with HSV-1 antigens, both alone and in combination with HERV antigens. (IL-2 and IL-10 production were unchanged.)¹⁰³</p> <ul style="list-style-type: none"> • Two studies^{104, 105} found serum or CSF immune complexes containing HSV antibodies and/or antigen in MS subjects (however, antibodies or antigens for other viruses were also detected).
<p><i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i></p>
<p>Not assessed</p>
<p><i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i></p>
<p>The effect of anti-herpes treatment on MS disease activity was examined by Lycke <i>et al.</i>,¹⁰⁶ who found that acyclovir reduced the number of exacerbations in RRMS subjects. Also, Bech <i>et al.</i>⁸⁴ found that valacyclovir appeared to reduce new active lesion formation in a subset of subjects who had active lesions at baseline.</p>
<p><i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i></p>
<p>A few relationships have been proposed involving retroviral activation or molecular mimicry:</p> <ul style="list-style-type: none"> • Brudek <i>et al.</i>¹⁰⁷ found a synergistic effect of HSV-1 (and HHV-6A) with HERV-H on cellular immune responses in both MS and control subjects, and in a subsequent study¹⁰⁸ showed that HSV-1 and other viruses induced higher reverse transcriptase activity in peripheral lymphocytes of MS subjects compared with controls • In addition, Perron <i>et al.</i>¹⁰⁹ detected increased activation of LM7 retrovirus in cells infected with HSV-1. • Cortese <i>et al.</i>¹¹⁰ and Esposito <i>et al.</i>⁸⁸ found antibodies cross-reactive for HSV-1 and brain proteins in MS sera and CSF.
<p><i>Conclusion:</i></p>
<p>The results of case/control studies do not strongly support a role for HSV in causing MS. However, antiviral treatment studies showing a reduction in MS lesion formation or relapses may suggest a role for HSV or other herpesviruses in stimulating disease activity.</p>

Pathogen: Human herpesvirus 6
<i>Description:</i>
Human herpesvirus 6 (HHV-6) is a common virus that infects most people in early childhood, most likely through salivary transmission. It is tropic for CD4 T cells but also has been shown to inhabit CNS tissue, and like many of the viruses described in this document has been detected in the CNS tissue and/or CSF of subjects with neurological conditions such as encephalitis. Two strains have been identified: HHV-6A and HHV-6B. The B strain is associated with the childhood disease exanthem subitum, also known as roseola; both strains have been found to be neurotropic. Like many other viruses, HHV-6 can be reactivated; its reactivation has been associated with several diseases.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
<p>Numerous studies have explored the question of whether HHV-6 is more prevalent in MS compared with non-MS subjects, or in plaque tissue vs. non-plaque tissue; other studies have focused on the presence of the two strains of HHV-6.</p> <ul style="list-style-type: none"> • HHV-6 has been detected in both MS brain tissue and control tissue, with some studies reporting that the virus is more prevalent in MS samples.^{28, 111, 112} • Some studies indicate a higher frequency or greater activity of HHV-6 in MS plaque tissue compared with normal appearing white matter (NAWM) or tissue from controls. For example, Challoner <i>et al</i>^β detected HHV-6 gene expression in the oligodendrocytes of MS subjects, particularly around plaques, but not in those of OND or NND controls. Goodman <i>et al</i>¹³ found HHV-6 DNA to be prevalent in the oligodendrocytes, lymphocytes, and microglia from biopsied acute MS lesions. Virtanen <i>et al</i>⁷ detected the presence of HHV-6 variant B antigen in 67% of MS lesional samples vs. 30% of normal control brain samples; HHV-6 antigen was located most often in oligodendrocytes, and frequently co-located with tissue plasminogen activator (tPA) protein in MS samples. A recent study of HHV-6 gene expression¹¹⁴ found higher levels of mRNA in MS brain samples (lesional samples, and to a lesser extent, NAWM) compared with normal control samples. On the other hand, Tuke <i>et al</i>¹¹⁵ found similar viral loads in lesion and non-lesion MS brain tissue samples, and Mameli <i>et al</i>¹¹⁶ could not detect HHV-6 replication in MS brain samples. • Studies of HHV-6 presence or activity in other tissues (blood, serum, PBMCs, and CSF) have provided mixed results; some studies detected no HHV-6 in MS or controls (e.g., Villoslada <i>et al</i>⁴⁵, Gutierrez <i>et al</i>¹⁷, and Riverol <i>et al</i>⁴⁶), others found similar frequencies of the virus in both MS and control samples (e.g., Ferrante <i>et al</i>²⁹), and others found a higher prevalence in MS compared with controls (e.g., Tejada-Simon <i>et al</i>¹⁸ and Alvarez-Lafuente <i>et al</i>⁴⁹). • In comparisons of strain A vs. strain B, some studies report detecting mostly HHV-6A in serum, urine or PBMCs, while others found mostly HHV-6B in serum or brain tissue. One study¹¹⁹ found only the B strain in PBMCs and only the A strain in cerebrospinal fluid or cells, whereas other studies found only variant A in PBMCs and serum of RRMS subjects with active HHV-6 replication⁴⁹, in serum from RRMS and SPMS subjects¹²⁰, and in CSF from RRMS subjects.³² In addition, HHV-6 DNA from the CSF of two natalizumab-treated MS subjects was subtyped as HHV-6A.¹²¹ Studies comparing the immune response to each strain (lymphoproliferative response, antibody analysis) in MS and control subjects offer mixed results (e.g., Ongradi <i>et al</i>¹²² and Soldan <i>et al</i>¹²³).

<ul style="list-style-type: none"> • A case report of a subject who developed Balo's concentric sclerosis describes a concurrent primary HHV-6 infection of the CNS which may have been related or may have been a coincidence¹²⁴.
<p><i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i></p>
<ul style="list-style-type: none"> • Antibody evidence from serum and CSF analysis has been mixed, although more studies (e.g., Villoslada <i>et al</i>⁴⁵, Caselli <i>et al</i>¹²⁵, and Virtanen <i>et al</i>¹²⁶) found higher titers or positivity in MS subjects vs. controls than found no differences between the groups. Associations with MS have been found for both IgG and IgM antibodies, as well as antibodies to various proteins (early and late antigens, U94/rep). • One study found a higher lymphoproliferation response to HHV-6 in MS cases than controls (to 6A only);¹²³ another found a similar response in both groups.¹²⁷ • Incubation of PBMCs with HHV-6 antigens did not result in an increased production of IFN-gamma in MS cases compared with controls, although an increased response was seen when combined HHV-6 and HERV antigens were used.¹⁰³ • One study showed a higher frequency in MS cases vs. controls of T cells cross-reactive to MBP and HHV-6 epitopes.¹²⁸ However, another study found that cross-reactive T cell lines were more readily generated in MS samples following MBP stimulation than HHV-6 stimulation and that the percentage of cross-reactive cell lines following HHV-6 stimulation was roughly equivalent in MS subjects and controls.¹²⁹
<p><i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i></p>
<p>Not assessed</p>
<p><i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i></p>
<p>Although the MS disease-modifying drug interferon-beta does not specifically target HHV-6, IFN-β has been found to reduce HHV-6 viral replication and perhaps exerts a beneficial effect in MS by this mechanism.¹³⁰⁻¹³² Although it is unclear whether to what extent the antiviral drug valacyclovir can inhibit HHV-6⁵⁴, two clinical trials of valacyclovir have been performed, using doses which may have some efficacy against HHV-6. Results from these trials indicate a possible clinical or MRI effect in subjects with severe or highly active MS.^{84, 85}</p>
<p><i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i></p>
<p>Several observations or hypotheses have been proposed and investigated to explain how HHV-6 might trigger MS:</p> <ul style="list-style-type: none"> • Infection of T cells with HHV-6 may be indirectly toxic to oligodendrocytes.¹³³ • HHV-6 and endogenous retroviruses may interact in a way that contributes to MS. HHV-6 and other herpesviruses have been shown to induce higher reverse transcriptase activity in blood cells of MS subjects compared with controls¹⁰⁸, and the combination of HHV-6 and HERV antigens has a synergistic effect on stimulating cellular immune responses.^{103, 107} • There is evidence, although mixed, for molecular mimicry based on increased T-cell reactivity (see above). • Elevated levels of soluble CD46, the receptor for HHV-6, have been found in the sera of MS cases compared with healthy and OND controls, particularly in MS cases positive for HHV-6 DNA in serum (but note that elevated soluble CD46 levels were also detected in the sera of subjects with other inflammatory

- diseases compared with healthy controls).¹³⁴
- As with other viruses such as EBV, reactivation may be associated with disease activity/exacerbations and progression.^{48, 49, 120, 132, 135-138}
 - Infection with HHV-6 has been shown *in vitro* to induce morphological changes in and inhibit proliferation of glial precursor cells¹³⁹; this effect may impair remyelination processes in MS or other diseases.
 - Chronic infection of astrocytes with HHV-6 impairs their glutamate uptake, which may lead to glutamate levels in the CNS that are harmful to oligodendrocytes.¹⁴⁰
 - HHV-6 encodes the protein U24, which shares a 7 amino acid stretch in common with an MBP phosphorylation site; MBP in MS subjects has lower levels of phosphorylation which may be a result of phospho-U24 confounding of signaling pathways.¹⁴¹

Conclusion:

It is possible that HHV-6 is involved in triggering MS. There is some evidence that HHV-6 is preferentially found in plaque tissue of MS subjects but the significance of this is unknown. Some studies have also identified differences in the antibody response to HHV-6 between MS subjects and controls. Many theories exist concerning how this virus may participate in the pathogenesis of MS (molecular mimicry, impairment of glutamate uptake in astrocytes, interaction with HERVs, infection of T cells, etc.).

Pathogen: Human immunodeficiency virus (HIV)
<i>Description:</i>
Human immunodeficiency virus (HIV), also referred to in older texts as HTLV-III, is the human retrovirus that causes AIDS (acquired immune deficiency syndrome). HIV infects CD4 T cells and macrophages, and AIDS is characterized by depletion of CD4 T cells which leads to opportunistic infections and tumors as well as dementia and other neurological abnormalities. HIV is chiefly transmitted through sexual contact, through contact with blood (e.g., transfusion of contaminated blood), and perinatally.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
Searches for HIV genetic sequences in MS CNS tissue and blood have produced negative results (e.g., Hauser <i>et al</i> ⁴² and Rozenberg <i>et al</i> ⁴³).
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
Several antibody analyses have been conducted using MS blood samples. Most found no reactivity to HIV/HTLV-III in MS subjects, although Ferrante <i>et al</i> ⁴⁰ did find a clear or weak antibody response to HIV in 10% of MS subjects, and Perron <i>et al</i> ⁴⁴ did find increased reactivity to reverse transcriptase fragments in MS cases compared with controls.
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
Not assessed
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
Not assessed
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
Not assessed
<i>Conclusion:</i>
No strong evidence exists for an association between MS and HIV. However, there is evidence for possible increased reactivity to retroviral components (see also the discussions of HTLV-1 and endogenous retroviruses).

Pathogen: Human T-cell leukemia (or T-lymphotropic) virus (HTLV)
<i>Description:</i>
<p>There are two known strains of HTLV: HTLV-1 and HTLV-2. HTLV-1 is an exogenous human retrovirus that is prevalent to varying degrees in different parts of the world and is particularly common in southwestern Japan and the Caribbean. Most HTLV-1 infected individuals have an asymptomatic infection, but a small percentage develop an associated disease such as adult T-cell leukemia/lymphoma. Approximately 0.25% of infected people develop HTLV-I-associated myelopathy/tropical spastic paraparesis (HAM/TSP), a demyelinating disease that resembles MS in many aspects. Transmission of the virus typically occurs through sexual contact, contact with contaminated blood, or breastfeeding. HTLV-2 is a closely related virus that has not yet been conclusively associated with any human disease.</p>
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
<p>HTLV-1 DNA has been detected in tissues of MS subjects by some studies but not by others:</p> <ul style="list-style-type: none"> • Although a few early studies associated MS with the presence of HTLV DNA sequences such as pol, env, and p24,^{145, 146} a large number of follow-on studies failed to find various HTLV DNA sequences in a variety of tissues (PBMCs, brain, CSF, etc.) in MS subjects (e.g., Watanabe <i>et al</i>,¹⁴⁷ Oksenberg <i>et al</i>,¹⁴⁸ and Ehrlich <i>et al</i>¹⁴⁹). • One study found increased detection of HTLV-1 tax-rex DNA (detected in 35.7% of cases) in MS PBMCs on day 10 of an acute attack; other viruses were also detected.⁴⁰
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
<p>Evidence is mixed:</p> <ul style="list-style-type: none"> • Approximately 40 studies have assessed the presence of HTLV-1 antibodies in MS subjects. Most found no evidence of seropositivity in MS subjects (e.g., French <i>et al</i>,¹⁵⁰ Merelli <i>et al</i>¹⁵¹), a few found equal rates in MS subjects vs. various types of controls (e.g., Kuroda <i>et al</i>,¹⁵² Brookes <i>et al</i>¹⁵³), and a handful found higher seropositivity in MS subjects vs. controls (e.g., Odum <i>et al</i>,¹⁵⁴ Ferrante <i>et al</i>¹⁵⁵). It has been pointed out that some cases of seroreactivity to HTLV antigens in MS subjects could be due to cross-reactivity with autoantibodies to endogenous retroviruses.¹⁵⁶ • No evidence has been produced of lymphocyte proliferation indicating HTLV-1 infection,¹⁵⁷ of cytopathic effects,¹⁵⁸ or of seronegative HTLV-1 infection.¹⁵⁹ One study did find that cells from MS subjects but not controls reacted to anti-p19 and anti-p24 antibodies.¹⁶⁰
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
Not assessed
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
Not assessed
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
Not assessed
<i>Conclusion:</i>
At this time the evidence does not indicate a causal role of HTLV-1 in MS. However,

the existence of a similar demyelinating disease (HAM/TSP) that is thought to be caused or triggered by HTLV-1 suggests that the involvement of retroviruses should continue to be studied in MS.

Pathogen: Human endogenous retroviruses (HERVs)
<i>Description:</i>
<p>HERVs are retrovirus-like elements that make up as much as 8% of the human genome and are found in both coding and non-coding regions. Most are unable to replicate because of genetic defects, but many HERVs are capable of encoding retroviral proteins. Expression of HERV RNA may be increased at sites of inflammation; HERVs may also be transactivated by herpesviruses¹⁰⁸. Several different HERVs have been investigated in MS. One, multiple sclerosis-related virus (MSRV, previously called LM7), is particularly noteworthy because it was first isolated from cells from people with MS. Following are brief research summaries for each of the individual HERVs investigated for involvement in MS.</p>
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
<ul style="list-style-type: none"> • <u>HRES-1</u>: One study found that expression levels of HRES-1 in brain tissue did not differ between MS and non-MS subjects.¹⁶¹ A few studies of HRES-1 genotypes have identified allelic differences between European MS and non-MS subjects, but no differences have been found in Chinese populations.¹⁶²⁻¹⁶⁵ • <u>ERV3</u>: In studies of ERV3 transcription, most identified no differences between MS and control groups, although one study¹⁶¹ showed that healthy controls had a higher frequency of an ERV3/zinc finger composite transcript. No allelic associations have been found for ERV3 and MS.^{166, 167} • <u>RGH/HERV-H/RTVL-H</u>: Three studies have reported an association with MS for the expression of RGH sequence or splice variants in plasma,¹⁶⁸⁻¹⁷⁰ however, another study using brain tissue found no HERV-H upregulation in MS.¹⁷¹ A study of CSF in MS and other neurological disease subjects found no samples positive for HERV-H.³² • <u>HERV-K</u>: One study reported that HERV-K expression was increased in brain tissue of subjects with MS or AIDS compared with healthy controls, as was expression of TNF-alpha;¹⁷² however, two other studies^{161, 171} found no increased expression of HERV-K in blood or brain tissue. Another study found equal expression of HML6 RNA in MS plaque versus control tissue.¹⁷³ With respect to HERV-K genotypes, an investigation of the HERV-K113 allele found that its presence was not significantly increased in MS subjects compared with their parents,¹⁷⁴ contradicting results from a previous, smaller study.¹⁷⁵ However, Tai, <i>et al</i>¹⁷⁶ did find a genetic association with MS for the K18.3 allele of HERV-K18 Env. • <u>MSRV/LM7/HERV-W/ERVWE1</u>: <ul style="list-style-type: none"> ○ Numerous studies have found the expression of HERV-W RNA to be increased in the blood, brain, or CSF samples of MS or optic neuritis subjects compared with other disease controls or healthy controls (e.g., Perron <i>et al</i>¹⁷⁷, Menard <i>et al</i>,¹⁷⁸ Serra <i>et al</i>,¹⁷⁹ Nowak <i>et al</i>¹⁸⁰, Mameli <i>et al</i>¹¹⁶). However, increased expression of MSRV RNA has also been shown in unaffected close relatives of MS subjects¹⁸¹, as well as in subjects with other diseases, particularly other inflammatory neurological diseases.¹⁸² Also, Petzold, <i>et al</i>¹⁸³ detected no evidence of MSRV viremia in acute optic neuritis subjects and Alvarez-Lafuente, <i>et al</i>⁹² found no MS or other neurological disease CSF samples to be positive for HERV-W sequences. ○ DNA levels of syncytin-1 (the Env protein of ERVWE-1) have been

detected at higher levels in MS vs. non-MS brain tissue.¹⁸⁴ In addition, a study comparing two members of the HERV-W family, MSR/V and ERVWE-1, found ERVWE-1 but not MSR/V DNA to be more abundant in MS brain tissue than control tissue; DNA levels for both ERVs were similar in MS and control PBMC samples. ERVWE-1 mRNA was also amplified in MS brain samples vs. control samples whereas no difference was seen for MSR/V¹⁸⁵.

- Expression of MSR/V/HERV-W has been associated with disease severity and/or duration of MS. For example, Dolei *et al*¹⁸² found MSR/V RNA in the CSF of 50% of MS subjects at onset, with the prevalence increasing with disease duration, and Mameli *et al* similarly found viral load to be directly related to disease duration.¹⁸⁶ Sotgiu, *et al*¹⁸⁷, associated MSR/V RNA in the CSF with faster progression, more relapses, and greater risk of conversion to a secondary progressive course. The presence of MSR/V RNA in blood and/or CSF of acute optic neuritis subjects was associated in another study (Sotgiu *et al*¹⁸⁸) with conversion to MS.
- Perron *et al*¹⁸⁹, using immunohistochemical methods, found HERV-W GAG and ENV expressed in both MS and normal brain, although the cells expressing these proteins differed between the two states. However, Mameli *et al* found MSR/V/HERV-W immunoreactivity within MS plaques (generally only active ones) but not within normal appearing white matter, perilesional areas, or tissue from normal controls.¹¹⁶
- Regarding genomic differences, one study of MSR/V *pol* sequence copy number found a greater number of sequences in the chromatin of MS subjects compared with controls.¹⁹⁰

Does immune response to pathogen differ between MS subjects and non-MS subjects?

- HRES-1: Elevated antibody titers were found in MS subjects compared with healthy controls, but titers were also elevated in other disease states such as Sjogren's.¹⁵³
- RGH/HERV-H/RTVL-H: Christensen *et al*¹⁷⁰ found increased antibody responses to HERV-H/RGH-2 in the serum and CSF of MS subjects compared with healthy controls and patients with other diseases. A follow-up study found that MS subjects, particularly those with active disease, had increased seroreactivity to HERV-H antigens compared with unaffected relatives; degree of reactivity (high or low) to HERV-H was often found to be similar among family members.¹⁹¹
- MSR/V: Stimulation of PBMCs by the surface protein ENV-SU was shown to induce a stronger production of cytokines IFN-gamma, IL-6 and IL-12p40 in MS subjects compared with controls.¹⁹² However, Ruprecht *et al* found serum antibodies against HERV-W Env (Syncytin-1) in only one of 50 MS subjects and no subjects positive for antibodies against MSR/V matrix, Gag and Env proteins; similarly, no cytotoxic T-lymphocyte responses were seen to various MSR/V/HERV-W-derived peptides.¹⁹³

Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?

Not assessed

Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?

Although interferon beta is not specifically targeted at inhibiting HERV activity, a small eleven-person trial did find that initiation of IFN-b therapy reduced MSR/V RNA blood levels in all subjects to below detection levels. These levels were maintained for a

year in ten of the subjects; MSRV levels rebounded in one subject, whose EDSS score also increased.¹⁸⁶

If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?

- RGH/HERV-H/RTVL-H: Combinations of HERV and herpesvirus (such as HHV-6A, HSV-1, and VZV) antigens have been shown to exert a synergistic effect on stimulating cellular immune responses in both MS subjects and controls.^{103, 107}
- LM7/MSRV/HERV-W: Antony *et al*¹⁷¹ showed that the HERV-W encoded protein syncytin is upregulated in glial cells in acute MS lesions, and that syncytin is cytotoxic to oligodendrocytes through the release of redox reactants. Previously, Menard *et al*¹⁷⁸ had reported that monocyte/macrophage culture supernatants were toxic to astrocytes and oligodendrocytes, and that this gliotoxicity was correlated with the reverse transcriptase activity in these cultures. In addition, Perron *et al*¹⁹⁴ demonstrated that MSRV envelope protein can induce an abnormal T-lymphocyte response and may act as a superantigen. This hypothesis was further explored in a transfer of MSRV-particles into SCID mice¹⁹⁵ and in a study of cytokine production in response to MSRV ENV-SU exposure.¹⁹² The latter study showed greater pro-inflammatory cytokine production in MS vs. control samples, which was correlated with disease severity.

Conclusion:

No conclusion can yet be drawn about the involvement of endogenous retroviruses in MS. The increase in MSRV expression activity in MS subjects, combined with its production of proteins that may be harmful to oligodendrocytes or act as superantigens, suggest a role for this virus. However, it should be noted that increased MSRV expression activity is not specific to MS but has also been seen in other diseases. Interestingly, two studies have identified familial patterns in seroreactivity to ERV antigens¹⁹¹ and expression of ERV sequences¹⁸¹; whether and how this might relate to familial patterns of MS is not known.

Pathogen: JC virus
<i>Description:</i>
JC virus (JCV) is a relatively common virus that is found worldwide, with prevalence varying according to geographic region. In some areas the majority of the adults studied have been found to be seropositive for JCV. Normally the virus establishes a latent, asymptomatic infection in the kidneys (and possibly other sites in the body), which is kept under control by the immune system. However, if the immune system is suppressed, JCV may travel to and/or become activated in the brain, where it can cause a demyelinating disease called progressive multifocal leukoencephalopathy (PML). Notably, PML has been diagnosed in a small number of MS patients receiving the MS drug Tysabri, causing a temporary withdrawal of the drug from the market and the development of programs for monitoring those receiving the drug.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
The available evidence does not demonstrate an increased prevalence of JCV in MS compared with non-MS subjects: <ul style="list-style-type: none"> • None of the searches for JCV DNA or antigens in MS brain tissue samples have produced a positive result.^{3, 196-198} • Of studies analyzing the presence of JCV DNA in CSF, one detected JCV in 9% of MS subjects studied but none of the controls¹⁹⁹, another detected it in 5% of early MS subjects but none of the controls,²⁰⁰ while another found similar prevalence of JCV DNA in the CSF of MS subjects compared with controls⁴⁴. Four studies, including a study of 1,869 natalizumab recipients, failed to detect JCV DNA in MS CSF samples.²⁰¹⁻²⁰⁴ • Several studies (e.g., Agostini <i>et al</i>²⁰¹) have analyzed the excretion of JCV in urine in MS subjects and controls, as a marker of JCV activation, but none report an association with MS. • The presence of JCV DNA in blood or serum has also been found not to differ between MS subjects and controls,^{40, 199, 204, 205} and does not appear to be increased by interferon beta treatment.^{205, 206}
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
One study assessed the synthesis of intrathecal antibodies to JCV in subjects with PML and other neurological diseases; oligoclonal antibodies to JCV were found in the CSF of one subject with MS, but these may have been part of a polyspecific reaction. ²⁰⁷ Du Pasquier, et al analyzed cytotoxic lymphocyte response to JCV and found it to be stronger in MS subjects than in normal controls. ²⁰⁸
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
Not assessed
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
Not assessed
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
Not assessed
<i>Conclusion:</i>
Although the similarities between PML and MS might suggest a causal role for JCV in MS, there is currently no strong evidence to support this idea.

Pathogen: Measles
<i>Description:</i>
The measles virus is a paramyxovirus that, in an unvaccinated person, can cause a disease involving rash, fever, runny nose, conjunctivitis and other complications. The disease is marked by a viremia that spreads to the lymphatic system, skin, respiratory system, intestines and urinary tract. The virus may also spread to the brain where it occasionally results in encephalitis. It can persist in the brain and at a later time reactivate to cause an inflammatory, demyelinating disease called subacute sclerosing panencephalitis (SSPE). Measles RNA can be detected in the brain tissue of people with SSPE. Note that an effective vaccine to measles became available in the US in 1963; since the implementation of public health vaccination programs, the incidence of measles in the US and other countries has been dramatically reduced.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
Mixed results have been provided by studies investigating the presence of measles antigens or RNA/DNA sequences in MS brain; some found the virus to be present in certain MS samples (e.g., Rastogi <i>et al</i> ²⁰⁹) while others failed to detect it (e.g., Dowling <i>et al</i> ²¹⁰). The most recent study of this type was Geeraedts <i>et al</i> ⁹ which did find evidence of a conserved morbillivirus protein in MS brain samples, but no evidence of specific measles proteins or mRNA using immunostaining and RT-PCR. Very few studies seem to have compared the presence of measles in MS vs. control brain. A small number of studies have searched for the presence of measles in other tissues such as the jejunum or peripheral blood lymphocytes (e.g., Woyciechowska <i>et al</i> , ²¹¹ Gupta <i>et al</i> , ²¹² Brankin <i>et al</i> ²¹³), but again their results are mixed.
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
This question has been analyzed extensively, particularly in studies of anti-measles antibodies which often report finding increased titers and/or prevalence in MS subjects compared with controls. <ul style="list-style-type: none"> • Approximately 50 studies (e.g., Fewster <i>et al</i>²¹⁴) have analyzed measles antibody titers in MS subjects vs. various types of controls (OND subjects, healthy controls, siblings and other family members, for example). Most of these studies have analyzed serum but occasionally have tested CSF or other substances such as tears. The majority of these studies found higher titers in MS subjects. Subjects with SSPE were often also tested and likewise found to have higher titers. • Several studies also found an increased prevalence of anti-measles antibodies in subjects with MS vs. controls. Evidence of intrathecal production of measles antibodies has been repeatedly found in MS subjects (e.g., Rostrom <i>et al</i>,²¹⁵ Schadlich <i>et al</i>,²¹⁶ Reiber <i>et al</i>²¹⁷). One study found measles antibodies in brain tissue from MS subjects, along with antibodies to other viruses,²¹⁸ however, another study found no measles antibodies in MS brain tissue.²¹⁹ • Many other studies have searched for measles-related immunological differences between MS subjects and controls (based on lymphocyte transformation, interferon response, lymphocyte-mediated cytotoxicity, and other assays). Although a few studies appeared to indicate a deficient response to measles virus (e.g., Neighbour <i>et al</i>,²²⁰ Jacobson <i>et al</i>²²¹), others reported similar responses in cases and controls (e.g., Tovell <i>et al</i>,¹³ Santoli <i>et al</i>²²²). • Some studies (e.g., Walker <i>et al</i>,²²³ Salonen <i>et al</i>,²²⁴ and Makela²²⁵) searched for correlations between immune responses and HLA status such as DR2 status. It

<p>is possible that the prevalence of the DR2 allele in MS may underlie some of the immunological responses to measles since class II genes have been found to influence responses to measles antigens (for instance, see Ovsyannikova <i>et al</i>²²⁶).</p>
<p><i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i></p>
<p>Approximately 17 studies have analyzed age of infection or other features of prior measles infections in people with MS vs. controls. Several showed that people with MS tended to have been infected later in life than controls (e.g., Haile <i>et al</i>,²²⁷ Sullivan <i>et al</i>,²²⁸ and Bachmann and Kesselring²²⁹); however, other studies showed that infection at earlier ages, such as before age 7, is associated with MS (e.g., Casetta <i>et al</i>²³⁰). Still others found no significant correlation between risk of MS and age of measles infection (e.g., Bager <i>et al</i>²³¹).</p>
<p><i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i></p>
<p>The dramatic reduction in measles cases starting in the 1960's due to vaccination may appear to rule out measles infection as a cause of MS (as is postulated in Svenningsson <i>et al</i>²³²). However, since the vaccine is live attenuated virus, there is a possibility that it could persist dormant somewhere in the body and later cause disease. It is also possible that physiological changes induced by the vaccine, such as a humoral or cellular immune response, could contribute to the etiology of MS.</p>
<p><i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i></p>
<p>Several investigators have looked into the possibility of immune cell cross-reactivity or molecular mimicry triggered by measles infection. Matossian-Rogers <i>et al</i>²³³ found T cell lines cross-reactive to MBP and measles virus in healthy controls, but in Richert <i>et al</i>²³⁴ and Pette <i>et al</i>,²³⁵ no measles-specific T cell lines from MS subjects or healthy controls showed any reactivity to MBP. Also, in Jingwu <i>et al</i>²³⁶ and in Rubio and Cuesta,²³⁷ no antibody cross-reactivity was found between MBP and measles virus.</p>
<p>Conclusion:</p>
<p>Higher prevalence and titers of anti-measles antibodies have been found in MS subjects vs. controls; however, similar findings have also been reported for other pathogens. Due to vaccination programs, measles infection has nearly been eliminated in many countries where MS prevalence rates are the highest (for example, the US). This would suggest that, were the measles virus an important trigger of MS, incidence rates should have begun to decline in these countries unless the vaccine itself can also trigger MS.</p>

Pathogen: Mumps
<i>Description:</i>
The mumps virus is a paramyxovirus that is found throughout the world and transmitted through close person-to-person contact. Mumps epidemics can occur where vaccination is not practiced. Distribution of the virus from the respiratory system through the bloodstream results in systemic infection involving target organs such as the salivary glands, testes, ovaries, and the pancreas. The central nervous system can also be affected through the development of mumps meningitis, or more rarely, mumps encephalitis. Approximately one-third of mumps infections are subclinical. A vaccine for mumps was licensed in the US in 1967; since then the incidence of reported clinical mumps infection in this country has declined by 99%.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
Only one study in this category was identified; ²³⁸ it failed to find mumps RNA in MS brain tissue.
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
Evidence is mixed: <ul style="list-style-type: none"> • A few studies of antibody titers showed elevated mumps titers in MS serum or CSF samples compared with those taken from controls (e.g., Berr <i>et al</i>²³⁹). Likewise, a few analyses of MS CSF samples found evidence of intrathecal antibody production, generally in a minority of MS subjects (e.g., Sindic <i>et al</i>²⁴⁰). • Studies involving leukocyte migration, lymphocyte blast transformation, rosette formation, T-cell reactivity and/or interferon production have produced mixed results, with some but not all indicating a reduced immune response in the case of MS subjects (e.g., Tovell <i>et al</i>,¹³ Ilonen <i>et al</i>²⁴¹). • A few studies (e.g., Salonen <i>et al</i>²²⁴) found immune response to be correlated either with the presence of exacerbations or with genetic (Dw2) status.
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
<ul style="list-style-type: none"> • Of the seven epidemiological studies that assessed the age of mumps infection in MS cases vs. controls, five (e.g., Hays²⁴²) found an older age of infection in MS cases, one found infection by age 7 to be associated with MS,²⁴³ and one found no correlation between MS and age of mumps infection.²³¹ • Most studies of past mumps infection have found MS subjects no more likely to have had mumps than controls (e.g., Bansil <i>et al</i>²⁴⁴).
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
As stated above, mumps vaccination has been practiced since the late 1960's but this has not had a demonstrated effect on MS incidence.
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
Not assessed
<i>Conclusion:</i>
There is no strong evidence to support involvement of the mumps virus in the development of MS. Later age of mumps infection was associated with MS in multiple studies, but this was often also found true for other types of infections in these same studies. Similarly, the immunological differences between MS and control subjects found concerning the mumps virus have also been reported for other pathogens.

Pathogen: Parainfluenza and related paramyxoviruses (6/94 virus, Sendai virus)
<i>Description:</i>
In addition to measles and mumps, other paramyxoviruses have been investigated for a role in MS. These include strains of the human parainfluenza virus (which cause primarily respiratory system symptoms), animal viruses such as Sendai, simian virus 5, Newcastle disease virus, and canine distemper virus (described separately in this document), and the 6/94 virus which was originally isolated from MS brain tissue.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
Evidence concerning the presence of these paramyxoviruses in MS is limited: <ul style="list-style-type: none"> • A few groups reported seeing filament-like inclusions in MS brain tissue that were suggestive of paramyxovirus (e.g., Tanaka <i>et al</i>²⁴⁵); however, other investigators identified them as normally occurring cellular products, such as byproducts of degeneration or postmortem autolysis.^{246,247} • The 6/94 virus was originally discovered through analysis of MS brain tissue samples,²⁴⁸ but little subsequent research appears to have been performed to determine its prevalence in MS or control brains.
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
<ul style="list-style-type: none"> • Prevalence of antibodies to these viruses has not been determined to be significantly increased in MS subjects vs. controls in either serum or CSF. A few studies of antibody titers to parainfluenza-1 or 6/94 found levels to be higher in MS subjects (e.g., Cernescu <i>et al</i>²⁴⁹), but others found no such increase (e.g., Whitaker <i>et al</i>²⁵⁰). One study determined SV5 antibodies to be present in a large percentage of MS CSF samples and that SV5 antigen was able to absorb oligoclonal bands,²⁵¹ however, subsequent studies were either not able to replicate OCB absorbance or found it to be a phenomenon not specific to MS.^{252, 253} • A few studies (e.g., Ciongoli <i>et al</i>²⁵⁴) found a decreased response to parainfluenza or 6/94 virus in MS subjects vs. controls using leukocyte migration or lymphoblast transformation studies. Leukocyte migration and interferon production studies using the Sendai virus did not reveal any difference between MS cases and controls.^{13, 255-257} Also, mixed results were reported by two studies of interferon alpha production using Newcastle disease virus.^{258, 259}
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
Not assessed
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
Not assessed
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
Not assessed
<i>Conclusion:</i>
The available evidence does not strongly support the involvement of these paramyxoviruses in MS. Follow-up investigations of indications connecting one of these viruses with MS have generally failed to replicate the findings or have shown them not to be specific to the particular virus or to MS.

Pathogen: Rubella (German measles)
<i>Description:</i>
Rubella is a togavirus that causes German measles, a disease characterized by rash and lymphadenopathy. Rubella infection can be transmitted transplacentally and can cause birth defects in a developing fetus. The incidence of rubella infection in the US has declined by approximately 99% since the introduction of the rubella vaccine in 1969.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
Two studies that searched for rubella genomic sequences in MS brain tissue failed to detect them. ^{7, 238}
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
<ul style="list-style-type: none"> • Several studies found evidence for intrathecal production of rubella antibodies, but often detected the production of antibodies to other viruses as well (e.g., Reiber <i>et al</i>,²¹⁷ Rostasy <i>et al</i>²⁶⁰). • Two studies found no impairment of rubella-induced inhibition of leukocyte migration in MS subjects,^{255, 261} however, two other studies found a decreased response in MS subjects using lymphocyte transformation²⁴¹ and interferon alpha production²²⁴ assays. • One additional study¹⁰⁴ found immune complexes containing rubella antibodies in around one-quarter of MS cases (other viruses were also found in these complexes).
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
A few studies found earlier development of German measles to be associated with MS (e.g., Casetta <i>et al</i> ²³⁰), others found later infection to be associated with MS (e.g., Compston <i>et al</i> ²⁶²), and yet another found no association with age of infection. ²³¹
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
As with other viruses such as measles and mumps, widespread vaccination has nearly eliminated the incidence of rubella in several countries without a corresponding reduction in risk of MS
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
Not assessed
<i>Conclusion:</i>
The available evidence does not strongly suggest a role for rubella in causing MS. Immune abnormalities involving rubella, such as intrathecal synthesis of anti-rubella antibodies, have also been detected for other viruses and may be a nonspecific feature of the disease.

Pathogen: Vaccinia
<i>Description:</i>
The vaccinia virus is a poxvirus that is used to vaccinate people against smallpox. Serious complications from vaccination are rare but can include CNS effects, most notably encephalitis. Routine smallpox vaccination was ended in the US in 1972, but the vaccinia virus is still used to prepare recombinant vaccines for other diseases.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
Not assessed
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
<ul style="list-style-type: none"> • There is evidence of an abnormal antibody response to vaccinia in people with MS, demonstrated both by higher titers in CSF and serum compared to different types of controls,²⁶³ and by a higher prevalence of CSF antibodies and signs of intrathecal production in MS subjects (e.g., Cremer <i>et al</i>²⁶⁴). • Regarding cellular immunity, one study showed a weaker lymphocyte transformation response in MS cases vs. healthy controls²⁶⁵ but two other studies found no impairment in migration inhibition or lymphocyte proliferation.^{266, 267}
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
Not assessed
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
Not assessed
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
Not assessed
<i>Conclusion:</i>
Although vaccinia antibodies have been detected more frequently and at higher levels in MS cases than controls, this is also true of other viruses. The fact that routine vaccinia vaccination was discontinued in 1972 but nonvaccinated people continue to be diagnosed with MS also points away from a significant causal role for this virus in MS.

Pathogen: Varicella zoster
<i>Description:</i>
Varicella zoster virus (VZV) is the cause of chickenpox and historically has been encountered by most susceptible individuals in childhood, although a vaccine has recently been introduced to prevent infection. In unvaccinated people, the initial infection develops and resolves over a period of a few weeks. The course of the infection includes initial replication in the oropharynx, dissemination throughout the body by viremia, and development of a characteristic rash. VZV may subsequently persist in the dorsal root ganglia. If reactivated, VZV can cause herpes zoster, also known as shingles; this mainly occurs in older adults.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
<ul style="list-style-type: none"> • A handful of studies have looked for VZV genetic sequences in brain tissue or blood; most found either no evidence for viral DNA or similar prevalence rates in MS cases and controls (e.g., Challoner <i>et al</i>^β, Martin <i>et al</i>^{β1}). • An analysis of CSF detected VZV DNA in 31.6% of MS subjects (43.5% of RRMS subjects) compared with 10.7% of subjects with other neurological diseases.⁴⁴ However, another study of CSF samples taken from MS subjects at their first demyelinating event and from neurological controls found no samples positive for VZV DNA.³² • One study²⁶⁸ found the presence of VZV DNA in blood cells only in MS subjects experiencing a relapse, but not in MS subjects in remission nor in controls. A follow-up study reported similar results, with VZV DNA detected in PBMCs of 95% of MS subjects in relapse, 17% of MS subjects in remission, and no controls.⁴⁸ Further study detected VZV DNA and viral particles identical to VZV in the CSF of MS subjects during relapse, which decreased and/or disappeared during remission.²⁶⁹
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
<ul style="list-style-type: none"> • Results have been varied on the subject of whether serum titers or seropositivity to VZV is increased in people with MS vs. controls (e.g., Sundstrom <i>et al</i>,⁵⁷ Myhr <i>et al</i>⁷⁰). However, the prevalence of CSF antibodies may be higher in MS subjects,²⁷¹ and multiple studies have found evidence of intrathecal production of antibodies to VZV, as well as to other viruses such as rubella and measles (e.g., Reiber <i>et al</i>,²¹⁷ Sindic <i>et al</i>⁴⁰, Bednarova <i>et al</i>⁷²). • One study of cellular immunity³⁵ found a lower interferon production response to VZV by lymphocytes of MS subjects compared with controls. However, Brudek <i>et al</i> detected an increased production of IFN-gamma in PBMCs of MS subjects compared with controls when stimulated with VZV antigens or VZV combined with HERV antigens.¹⁰³
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
One study describes a north-south diminishing gradient of varicella infection that resembles that which has been found for MS. ²⁷³ Another study identified six MS subjects out of 82 who had varicella or zoster concurrent with the development or progression of MS. ²⁷⁴
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
The effect of anti-herpes treatment on MS disease activity was examined by Bech <i>et al</i> , ⁸⁴ who found that valacyclovir appeared to reduce new active lesion formation in a

subset of subjects who had active lesions at baseline. Another study ²⁷⁵ found improvement in 28% of MS subjects who were administered an attenuated varicella virus vaccine, but this was a short-term uncontrolled trial.
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
A recent study (Brudek <i>et al</i> ¹⁰⁸) determined that VZV antigens were capable of inducing reverse transcriptase expression from lymphocytes and that this effect was sustained for longer (> 6 days) than could be achieved with other viral antigens.
<i>Conclusion:</i>
A few associations between MS and VZV have been reported, such as a notably increased prevalence of DNA in MS relapses and presence of VZV antibodies in MS CSF samples. However, at this time, no conclusions can be drawn about any potential role of VZV in the etiology of MS.

III. Bacteria

Pathogen: <i>Borrelia burgdorferi</i>
<i>Description:</i>
Borrelia burgdorferi is a Gram-negative spirochete that causes Lyme disease, a disease that can affect multiple organ systems including the central nervous system. In fact, the neurological symptoms of chronic Lyme disease can closely resemble those of MS, which creates challenges in the differential diagnosis of these diseases. Diagnosis is also complicated by the possibility of false positives and false negatives in detecting antibody responses to B. burgdorferi which is the standard diagnostic approach for Lyme disease. In the US, B. burgdorferi is transmitted by the bite of hard ticks such as the deer tick in the northern and midwestern US and the western black-legged tick in western states. Lyme disease is endemic to those regions inhabited by these ticks, notably wooded areas in the northeast US, the Great Lakes region, and the Pacific Northwest.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
Not assessed
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
Evidence is inconclusive – several studies have measured the prevalence of antibodies to B. burgdorferi in MS and control subjects, but the results have varied greatly. In some studies, antibodies were found at low frequencies in MS subjects and/or less often than in controls (e.g., Schmutzhard <i>et al</i> ²⁷⁶). In other studies, antibodies were found in a high percentage of MS subjects (for instance, 38.5% according to one study ²⁷⁷) and more frequently than in controls. Differences in antibody positivity rates between studies may be influenced by overall Lyme prevalence in the geographic regions where these studies were performed and/or differences in lab methodologies.
<i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i>
Not assessed
<i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i>
Not assessed
<i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i>
Some studies have found evidence for cross-reactivity between borrelia proteins and neural antigens such as MBP (e.g., Weigelt <i>et al</i> ²⁷⁸).
<i>Conclusion:</i>
Because it is endemic only to certain geographic locations, B. burgdorferi is unlikely to be an MS trigger in all cases. However, assuming that MS is an etiologically heterogeneous disease, in individuals who are predisposed to MS, it is plausible that B. burgdorferi infection may initiate certain pathogenic events leading to MS (see Karussis <i>et al</i> ²⁷⁹ which discusses an apparent case of concomitant Lyme disease and MS).

Pathogen: Chlamydia pneumoniae
<i>Description:</i>
Chlamydia pneumoniae (Cpn) is a common obligate intracellular bacteria that causes respiratory tract infections and is transmitted by respiratory secretions. It has been implicated as a risk factor in other diseases, most notably heart disease, but also other neurological disorders such as Alzheimer's disease.
<i>Is presence of actual pathogen different in MS subjects compared with non-MS subjects?</i>
<p>Some positive findings have been reported, but overall the evidence is inconclusive:</p> <ul style="list-style-type: none"> • Four studies have searched for the presence of Cpn DNA in MS brain tissue; three reportedly failed to detect any DNA²⁸⁰⁻²⁸² but the fourth detected MOMP and 16s RNA DNA in 5 of 8 MS subjects and 3 of 18 OND controls²⁸³. This last study also performed immunohistochemical staining of brain tissue, and found evidence of staining with three anti-chlamydial antibodies in 7 of 20 MS subjects vs. 0 of 17 OND subjects. (Staining was confined to ependymal surfaces and periventricular regions.) • Over 20 studies have analyzed the presence of Cpn DNA in MS CSF, with or without an OND control group. Several failed to detect DNA in any of the samples (e.g., Saiz <i>et al.</i>,²⁸⁴ Derfuss <i>et al.</i>²⁸⁵, Budak <i>et al.</i>²⁸⁶, Lindsey and Patel²⁸⁷); however, several others were able to detect it in at least some samples. Of those studies that detected DNA and compared its prevalence in MS and controls, some found it to be more common or present only in MS subjects (e.g., Sotgiu <i>et al.</i>,²⁸⁸ Hao <i>et al.</i>²⁸⁹), while the rest found it to be equally prevalent in MS subjects and controls (e.g., Contini <i>et al.</i>²⁹⁰). These discrepancies in CSF PCR studies may be due to a number of factors such as variations in the techniques used; currently no standardized process exists (see Yamamoto²⁹¹ for a discussion). • One study using electron microscopy detected structures resembling chlamydial bodies in the CSF of 11/20 MS subjects and 2/12 OND controls²⁸³. • Studies that have analyzed blood or serum samples have provided mixed results; a recent analysis of PBMCs from 112 MS subjects using real-time PCR detected Cpn DNA in only two subjects, and in these subjects the viral loads were low²⁹².
<i>Does immune response to pathogen differ between MS subjects and non-MS subjects?</i>
<ul style="list-style-type: none"> • Several studies have analyzed the prevalence of antibodies to Cpn in MS subjects and controls, usually OND controls, but occasionally healthy controls. In general, these studies have found serum positivity to be equivalent in MS subjects and controls, but results from studies analyzing CSF positivity or intrathecal synthesis were mixed, with some showing higher frequencies in MS subjects (e.g., Krametter <i>et al.</i>²⁹³) and others showing similar frequencies in MS and OND control groups (e.g., Sotgiu <i>et al.</i>²⁸⁸). A study that found greater evidence of active Cpn infection in MS cases than controls (see Parratt <i>et al.</i> below) found serum Cpn-specific IgA to be more prevalent in healthy controls, indicating that IgA is not a reliable indicator of current infection.²⁹⁴ • Antibody titers in CSF have generally been found to be higher in MS subjects vs. controls, usually OND controls (e.g., Sriram <i>et al.</i>²⁹⁵), but serum titers have often been found to be similar in MS subjects and OND or healthy controls (e.g., Villoslada <i>et al.</i>⁴⁵, Krametter <i>et al.</i>²⁹⁶). • One prospective study using serum samples from two cohorts (US Army and Kaiser Permanente) found no association in either cohort between Cpn

<p>seropositivity and risk of MS, but did report an association between serum IgG anti-Cpn levels and MS risk in the Kaiser Permanente cohort²⁹⁷.</p> <ul style="list-style-type: none"> • Serum Cpn-specific immune complexes, a sign of active infection, were found by Parratt <i>et al</i> in 24% of MS subjects compared with 16% of other neurological disease controls and 15% of healthy controls (4.7% when residents of a certain town were excluded). The presence of Cpn-specific immune complexes was not associated with MS disease duration, and is therefore probably not simply a function of disease burden. 30%-54% of the subjects who were positive for immune complexes were negative for IgG and IgA antibodies to Cpn.²⁹⁴ • Yao <i>et al</i>²⁹⁸ found oligoclonal bands from MS subjects to be partially or completely adsorbed by Cpn antigens but not by a variety of other antigens. However, Derfuss <i>et al</i>²⁸⁵ reported a lack of reactivity of oligoclonal bands to Cpn elementary or reticulate bodies.
<p><i>Does age of infection or other characteristic of infection differ significantly between MS subjects and non-MS subjects?</i></p>
<p>Not assessed</p>
<p><i>Have any therapies aimed at controlling or eliminating the pathogen been found to alter the risk or clinical features of MS?</i></p>
<p>Sriram <i>et al</i>²⁹⁹ presents a case report of a man with MS who was positive for Cpn DNA and antibodies in his CSF and who experienced improvement of his MS symptoms following anti-chlamydial treatment. A small (8 subject, 14 month) follow-on study of azithromycin and rifampin showed no significant treatment effect on lesion volume or number, but reported a significant increase in brain atrophy in the placebo group³⁰⁰. However, a separate study (Woessner <i>et al</i>³⁰¹) found no significant therapeutic effect of a different regimen consisting of three six-week cycles of roxithromycin. An analysis of medical records of MS cases in the three years prior to MS onset and matched controls did not find a protective effect for antibiotics active against Cpn, but did detect a lower risk for subjects treated with penicillin³⁰².</p>
<p><i>If any significant differences appear to exist for any of the above, have cause-effect relationships been established to rule out other types of connections?</i></p>
<ul style="list-style-type: none"> • MacIntyre <i>et al</i>³⁰³ suggests that Cpn infection of human brain microvascular endothelial cells may alter the permeability of the blood-brain barrier. • Lenz <i>et al</i>³⁰⁴ reports on potential mimicry between an MBP peptide and a Cpn peptide that can induce EAE in rats. • Four studies found Cpn DNA positivity in MS to be associated with MRI and/or clinical activity^{289, 290, 305, 306}. Another found Cpn infections to be associated with risk of MS exacerbation.³⁰⁷ Whether these findings derive from a causal role of Cpn in the pathogenesis of MS has yet to be determined.
<p><i>Conclusion:</i></p>
<p>The available evidence concerning Cpn and MS is quite mixed. Although some studies have linked the presence of Cpn DNA or antibodies in CSF to MS, not all have shown such a connection. Furthermore, the one prospective study conducted²⁹⁷ to analyze Cpn antibodies prior to MS onset showed only a possible association between IgG levels and risk of MS. However, the presence of chlamydial bodies in the CSF of MS subjects and the higher prevalence of anti-Cpn immune complexes in MS subjects support the possible involvement of this bacterium in MS. Investigations of anti-chlamydial treatments in MS have been limited and have evaluated different treatment regimens.</p>

IV. Other pathogens investigated in MS

In addition to the pathogens presented in the templates above, a number of other pathogens have been evaluated for an etiological role in MS but to a lesser degree. For each of the pathogens listed below, only a few studies (often only one or two) could be found that evaluated experimental evidence for its involvement in MS. Interestingly, a few of these infectious agents (such as hepatitis B virus and helminths) have been proposed to have a protective effect against MS, perhaps by mediating a shift in the T cell repertoire towards a Th2 profile.

Our infectious agents studies spreadsheet (which can be downloaded at www.acceleratedcure.org/downloads/phase2-infectiousagents-studies.xls) contains a listing of the studies found to date for each of these pathogens.

Viruses:

- Acute human encephalomyelitis virus
- Adult T-cell leukemia (ATL) virus
- BK virus
- Borna disease virus
- Bovine leukemia virus
- Caprine arthritis encephalitis virus
- Enteroviruses
- Hepatitis B
- Hepatitis C
- Hepatitis G
- Human herpesvirus 1
- Human herpesvirus 2
- Human herpesvirus 7
- Human herpesvirus 8
- Human herpesvirus 7
- Human herpesvirus 8
- Human papilloma virus
- Human parvovirus B19
- Inoue-Melnick virus
- Influenza virus
- Lipovnik virus
- Maedi-Visna virus
- Marek's disease virus
- Parvovirus B19
- Poliovirus
- Respiratory syncytial virus
- Simian immunodeficiency virus
- Simian virus 40

- Spumavirus/spumaretrovirus
- Torque Teno virus
- Vesicular stomatitis virus

Bacteria:

- Acinetobacter
- Bacillus Calmette-Guerin
- Bifidobacterium
- Bordetella pertussis
- Clostridium tetani
- Corynebacterium diphtheriae
- Enterococcus
- Escherichia coli
- Helicobacter pylori
- Hemophilus influenzae
- Microsporidia
- Mycobacteria bovis
- Mycobacteria leprae
- Mycobacteria tuberculosis
- Mycoplasma pneumoniae
- Parachlamydia-like organisms
- Pseudomonas aeruginosa
- Staphylococcus/S. aureus
- Streptococcus

Fungi:

- Candida albicans

Protozoa:

- Plasmodium falciparum
- Toxoplasma gondii

Helminths:

- Toxoplasma gondii
- Trichuris trichiura

V. Conclusion

The information presented in the templates above show that although many different infectious agents have been studied, some quite extensively, for a potential role in MS, currently no specific pathogen can be conclusively labeled as an MS trigger. There are many challenges inherent in identifying the triggers of a disease which may take many years to manifest itself and which may require

the contributions of multiple etiological factors. Assuming that MS is a multifactorial and heterogeneous disease, it is likely that the evidence implicating infectious triggers of MS will not be clear cut, will be opposed by conflicting findings, and will require careful interpretation. It is also possible that identifying and confirming infections triggers of MS will require the use of new investigative techniques and expanding the scope of research to pathogens not previously evaluated in MS.

References

1. Moore, F.G. and C. Wolfson, *Human herpes virus 6 and multiple sclerosis*. Acta Neurol Scand, 2002. **106**(2): p. 63-83. PubMed ID: 12100366.
2. Kirk, J. and A.L. Zhou, *Viral infection at the blood-brain barrier in multiple sclerosis:--an ultrastructural study of tissues from a UK Regional Brain Bank*. Mult Scler, 1996. **1**(4): p. 242-52. PubMed ID: 9345442.
3. Challoner, P.B., et al., *Plaque-associated expression of human herpesvirus 6 in multiple sclerosis*. Proc Natl Acad Sci U S A, 1995. **92**(16): p. 7440-4. PubMed ID: 7638210.
4. Appel, M.J., et al., *Canine viruses and multiple sclerosis*. Neurology, 1981. **31**(8): p. 944-9. PubMed ID: 6267515.
5. Andersen, O., et al., *Viral infections trigger multiple sclerosis relapses: a prospective seroepidemiological study*. J Neurol, 1993. **240**(7): p. 417-22. PubMed ID: 8410082.
6. Cook, R.D., R.L. Flower, and N.S. Dutton, *Light and electron microscopical studies of the immunoperoxidase staining of multiple sclerosis plaques using antisera to a feline-derived agent and to galactocerebroside*. Neuropathol Appl Neurobiol, 1986. **12**(1): p. 63-79. PubMed ID: 3010153.
7. Cosby, S.L., et al., *Examination of eight cases of multiple sclerosis and 56 neurological and non-neurological controls for genomic sequences of measles virus, canine distemper virus, simian virus 5 and rubella virus*. J Gen Virol, 1989. **70**(Pt 8): p. 2027-36. PubMed ID: 2769228.
8. Lassmann, H., et al., *A new paraclinical CSF marker for hypoxia-like tissue damage in multiple sclerosis lesions*. Brain, 2003. **126**(Pt 6): p. 1347-57. PubMed ID: 12764056.
9. Geeraedts, F., et al., *Search for morbillivirus proteins in multiple sclerosis brain tissue*. Neuroreport, 2004. **15**(1): p. 27-32. PubMed ID: 15106826.
10. Cook, S.D., P.C. Dowling, and W.C. Russell, *Neutralizing antibodies to canine distemper and measles virus in multiple sclerosis*. J Neurol Sci, 1979. **41**(1): p. 61-70. PubMed ID: 438844.
11. Kurtzke, J.F., et al., *Multiple sclerosis in the Faroe Islands. IV. The lack of a relationship between canine distemper and the epidemics of MS*. Acta Neurol Scand, 1988. **78**(6): p. 484-500. PubMed ID: 3223236.
12. Krakowka, S., et al., *Antibody responses to measles virus and canine distemper virus in multiple sclerosis*. Ann Neurol, 1983. **14**(5): p. 533-8. PubMed ID: 6197006.
13. Tovell, D.R., et al., *Interferon production by lymphocytes from multiple sclerosis and non-MS patients*. Neurology, 1983. **33**(5): p. 640-3. PubMed ID: 6188993.
14. Hodge, M.J. and C. Wolfson, *Canine distemper virus and multiple sclerosis*. Neurology, 1997. **49**(2 Suppl 2): p. S62-9. PubMed ID: 9270694.
15. De Keyser, J., et al., *Disappearance of beta2-adrenergic receptors on astrocytes in canine distemper encephalitis: possible implications for the pathogenesis of multiple sclerosis*. Neuroreport, 2001. **12**(2): p. 191-4. PubMed ID: 11209919.
16. Tanaka, R., Y. Iwasaki, and H. Koprowski, *Intracisternal virus-like particles in brain of a multiple sclerosis patient*. J Neurol Sci, 1976. **28**(1): p. 121-6. PubMed ID: 932771.
17. Burks, J.S., et al., *Two coronaviruses isolated from central nervous system tissue of two multiple sclerosis patients*. Science, 1980. **209**(4459): p. 933-4. PubMed ID: 7403860.

18. Stewart, J.N., S. Mounir, and P.J. Talbot, *Human coronavirus gene expression in the brains of multiple sclerosis patients*. *Virology*, 1992. **191**(1): p. 502-5. PubMed ID: 1413524.
19. Arbour, N., et al., *Neuroinvasion by human respiratory coronaviruses*. *J Virol*, 2000. **74**(19): p. 8913-21. PubMed ID: 10982334.
20. Dessau, R.B., G. Lisby, and J.L. Frederiksen, *Coronaviruses in brain tissue from patients with multiple sclerosis*. *Acta Neuropathol (Berl)*, 2001. **101**(6): p. 601-4. PubMed ID: 11515789.
21. Salmi, A., et al., *Antibodies to coronaviruses OC43 and 229E in multiple sclerosis patients*. *Neurology*, 1982. **32**(3): p. 292-5. PubMed ID: 6278362.
22. Leinikki, P., et al., *Virus antibodies in the cerebrospinal fluid of multiple sclerosis patients detected with ELISA tests*. *J Neurol Sci*, 1982. **57**(2-3): p. 249-55. PubMed ID: 6298370.
23. Fleming, J.O., et al., *Antigenic assessment of coronaviruses isolated from patients with multiple sclerosis*. *Arch Neurol*, 1988. **45**(6): p. 629-33. PubMed ID: 2835952.
24. Talbot, P.J., et al., *Myelin basic protein and human coronavirus 229E cross-reactive T cells in multiple sclerosis*. *Ann Neurol*, 1996. **39**(2): p. 233-40. PubMed ID: 8967755.
25. Edwards, J.A., F. Denis, and P.J. Talbot, *Activation of glial cells by human coronavirus OC43 infection*. *J Neuroimmunol*, 2000. **108**(1-2): p. 73-81. PubMed ID: 10900340.
26. Aulakh, G.S., P. Albrecht, and W.W. Tourtellotte, *Search for cytomegalovirus and herpes simplex virus genetic information in multiple sclerosis*. *Neurology*, 1980. **30**(5): p. 530-2. PubMed ID: 6245390.
27. Virtanen, J.O., et al., *Co-localization of human herpes virus 6 and tissue plasminogen activator in multiple sclerosis brain tissue*. *Med Sci Monit.*, 2005. **11**(3): p. BR84-7. PubMed ID: 15735559.
28. Sanders, V.J., et al., *Detection of herpesviridae in postmortem multiple sclerosis brain tissue and controls by polymerase chain reaction*. *J Neurovirol*, 1996. **2**(4): p. 249-58. PubMed ID: 8799216.
29. Ferrante, P., et al., *Molecular evidences for a role of HSV-1 in multiple sclerosis clinical acute attack*. *J Neurovirol*, 2000. **6**(Suppl 2): p. S109-14. PubMed ID: 10871797.
30. Alvarez-Lafuente, R., et al., *Prevalence of herpesvirus DNA in MS patients and healthy blood donors*. *Acta Neurol Scand*, 2002. **105**(2): p. 95-9. PubMed ID: 11903118.
31. Martin, C., et al., *Absence of seven human herpesviruses, including HHV-6, by polymerase chain reaction in CSF and blood from patients with multiple sclerosis and optic neuritis*. *Acta Neurol Scand*, 1997. **95**(5): p. 280-3. PubMed ID: 9188902.
32. Alvarez-Lafuente, R., et al., *Herpesviruses and human endogenous retroviral sequences in the cerebrospinal fluid of multiple sclerosis patients*. *Mult Scler.*, 2008. **14**(5): p. 595-601. PubMed ID: 18566025.
33. Ascherio, A., et al., *Epstein-Barr virus antibodies and risk of multiple sclerosis: a prospective study*. *Jama*, 2001. **286**(24): p. 3083-8. PubMed ID: 11754673.
34. Alotaibi, S., et al., *Epstein-Barr virus in pediatric multiple sclerosis*. *Jama*, 2004. **291**(15): p. 1875-9. PubMed ID: 15100207.
35. Haahr, S., A. Moller-Larsen, and E. Pedersen, *Immunological parameters in multiple sclerosis patients with special reference to the herpes virus group*. *Clin Exp Immunol*, 1983. **51**(2): p. 197-206. PubMed ID: 6188564.

36. Hollsberg, P., H.J. Hansen, and S. Haahr, *Altered CD8+ T cell responses to selected Epstein-Barr virus immunodominant epitopes in patients with multiple sclerosis*. Clin Exp Immunol, 2003. **132**(1): p. 137-43. PubMed ID: 12653848.
37. Lunemann, J.D., et al., *Increased frequency and broadened specificity of latent EBV nuclear antigen-1-specific T cells in multiple sclerosis*. Brain., 2006. **129**(Pt 6): p. 1493-506. Epub 2006 Mar 28. PubMed ID: 16569670.
38. Scotet, E., et al., *Frequent enrichment for CD8 T cells reactive against common herpes viruses in chronic inflammatory lesions: towards a reassessment of the physiopathological significance of T cell clonal expansions found in autoimmune inflammatory processes*. Eur J Immunol, 1999. **29**(3): p. 973-85. PubMed ID: 10092102.
39. Zivadinov, R., et al., *Positivity of cytomegalovirus antibodies predicts a better clinical and radiological outcome in multiple sclerosis patients*. Neurol Res., 2006. **28**(3): p. 262-9. PubMed ID: 16687051.
40. Ferrante, P., et al., *Human T-cell lymphotropic virus tax and Epstein-Barr virus DNA in peripheral blood of multiple sclerosis patients during acute attack*. Acta Neurol Scand Suppl, 1997. **169**: p. 79-85. PubMed ID: 9174643.
41. Hilton, D.A., et al., *Absence of Epstein-Barr virus RNA in multiple sclerosis as assessed by in situ hybridisation*. J Neurol Neurosurg Psychiatry, 1994. **57**(8): p. 975-6. PubMed ID: 7520057.
42. Morre, S.A., et al., *Is Epstein-Barr virus present in the CNS of patients with MS?* Neurology, 2001. **56**(5): p. 692. PubMed ID: 11245733.
43. Serafini, B., et al., *Dysregulated Epstein-Barr virus infection in the multiple sclerosis brain*. J Exp Med., 2007. **204**(12): p. 2899-912. Epub 2007 Nov 5. PubMed ID: 17984305.
44. Mancuso, R., et al., *Increased prevalence of varicella zoster virus DNA in cerebrospinal fluid from patients with multiple sclerosis*. J Med Virol., 2007. **79**(2): p. 192-9. PubMed ID: 17177306.
45. Villoslada, P., et al., *The immune response against herpesvirus is more prominent in the early stages of MS*. Neurology, 2003. **60**(12): p. 1944-8. PubMed ID: 12821737.
46. Riverol, M., et al., *Antibodies against Epstein-Barr virus and herpesvirus type 6 are associated with the early phases of Multiple Sclerosis*. J Neuroimmunol, 2007. **13**: p. 13 PubMed ID: 17869349.
47. Hay, K.A. and R.B. Tenser, *Leukotropic herpesviruses in multiple sclerosis*. Mult Scler, 2000. **6**(2): p. 66-8. PubMed ID: 10773849.
48. Sotelo, J., G. Ordonez, and B. Pineda, *Varicella-zoster virus at relapses of multiple sclerosis*. J Neurol., 2007. **254**(4): p. 493-500. Epub 2007 Mar 31. PubMed ID: 17401519.
49. Alvarez-Lafuente, R., et al., *Human herpesvirus 6 and multiple sclerosis: a one-year follow-up study*. Brain Pathol., 2006. **16**(1): p. 20-7. PubMed ID: 16612979.
50. Wagner, H.J., K.L. Munger, and A. Ascherio, *Plasma viral load of Epstein-Barr virus and risk of multiple sclerosis*. Eur J Neurol, 2004. **11**(12): p. 833-4. PubMed ID: 15667414.
51. Wandinger, K., et al., *Association between clinical disease activity and Epstein-Barr virus reactivation in MS*. Neurology, 2000. **55**(2): p. 178-84. PubMed ID: 10908887.
52. Torkildsen, O., et al., *Epstein-Barr virus reactivation and multiple sclerosis*. Eur J Neurol., 2008. **15**(1): p. 106-8. Epub 2007 Nov 27. PubMed ID: 18042233.

53. Fraser, K.B., et al., *Increased tendency to spontaneous in-vitro lymphocyte transformation in clinically active multiple sclerosis*. Lancet, 1979. **2**(8145): p. 175-6. PubMed ID: 90801.
54. Hollsberg, P., et al., *Presence of Epstein-Barr virus and human herpesvirus 6B DNA in multiple sclerosis patients: associations with disease activity*. Acta Neurol Scand., 2005. **112**(6): p. 395-402. PubMed ID: 16281923.
55. Banwell, B., et al., *Clinical features and viral serologies in children with multiple sclerosis: a multinational observational study*. Lancet Neurol., 2007. **6**(9): p. 773-81. PubMed ID: 17689148.
56. Levin, L.I., et al., *Multiple sclerosis and Epstein-Barr virus*. Jama, 2003. **289**(12): p. 1533-6. PubMed ID: 12672770.
57. Sundstrom, P., et al., *An altered immune response to Epstein-Barr virus in multiple sclerosis: a prospective study*. Neurology, 2004. **62**(12): p. 2277-82. PubMed ID: 15210894.
58. Levin, L.I., et al., *Temporal relationship between elevation of epstein-barr virus antibody titers and initial onset of neurological symptoms in multiple sclerosis*. Jama., 2005. **293**(20): p. 2496-500. PubMed ID: 15914750.
59. DeLorenze, G.N., et al., *Epstein-Barr virus and multiple sclerosis: evidence of association from a prospective study with long-term follow-up*. Arch Neurol., 2006. **63**(6): p. 839-44. Epub 2006 Apr 10. PubMed ID: 16606758.
60. Haahr, S., et al., *A role of late Epstein-Barr virus infection in multiple sclerosis*. Acta Neurol Scand, 2004. **109**(4): p. 270-5. PubMed ID: 15016009.
61. Sundstrom, P., et al., *EBNA-1 reactivity and HLA DRB1*1501 as statistically independent risk factors for multiple sclerosis: a case-control study*. Mult Scler., 2008. **14**(8): p. 1120-2. Epub 2008 Jun 23. PubMed ID: 18573815.
62. De Jager, P.L., et al., *Integrating risk factors: HLA-DRB1*1501 and Epstein-Barr virus in multiple sclerosis*. Neurology., 2008. **70**(13 Pt 2): p. 1113-8. Epub 2008 Feb 13. PubMed ID: 18272866.
63. Rand, K.H., et al., *Epstein-Barr virus nuclear antigen-1 (EBNA-1) associated oligoclonal bands in patients with multiple sclerosis*. J Neurol Sci, 2000. **173**(1): p. 32-9. PubMed ID: 10675577.
64. Cepok, S., et al., *Identification of Epstein-Barr virus proteins as putative targets of the immune response in multiple sclerosis*. J Clin Invest, 2005. **115**(5): p. 1352-60. Epub 2005 Apr 14. PubMed ID: 15841210.
65. Craig, J.C., et al., *Immunological control of Epstein-Barr virus-transformed lymphocytes in multiple sclerosis*. Clin Immunol Immunopathol, 1983. **29**(1): p. 86-93. PubMed ID: 6309449.
66. Craig, J.C., et al., *Subsets of T lymphocytes in relation to T lymphocyte function in multiple sclerosis*. Clin Exp Immunol, 1985. **61**(3): p. 548-55. PubMed ID: 3000660.
67. Craig, J.C., M. Haire, and J.D. Merrett, *T-cell-mediated suppression of Epstein-Barr virus-induced B lymphocyte activation in multiple sclerosis*. Clin Immunol Immunopathol, 1988. **48**(3): p. 253-60. PubMed ID: 2841052.
68. Lunemann, J.D., et al., *EBNA1-specific T cells from patients with multiple sclerosis cross react with myelin antigens and co-produce IFN-gamma and IL-2*. J Exp Med., 2008. **205**(8): p. 1763-73. Epub 2008 Jul 28. PubMed ID: 18663124.
69. Jilek, S., et al., *Strong EBV-specific CD8+ T-cell response in patients with early multiple sclerosis*. Brain., 2008. **131**(Pt 7): p. 1712-21. Epub 2008 Jun 11. PubMed ID: 18550621.

70. Gronen, F., et al., *Frequency analysis of HLA-B7-restricted Epstein-Barr virus-specific cytotoxic T lymphocytes in patients with multiple sclerosis and healthy controls*. J Neuroimmunol., 2006. **180**(1-2): p. 185-92. Epub 2006 Oct 4. PubMed ID: 17023054.
71. Operskalski, E.A., et al., *A case-control study of multiple sclerosis*. Neurology, 1989. **39**(6): p. 825-9. PubMed ID: 2725877.
72. Lindberg, C., et al., *Epidemiological investigation of the association between infectious mononucleosis and multiple sclerosis*. Neuroepidemiology, 1991. **10**(2): p. 62-5. PubMed ID: 2062419.
73. Martyn, C.N., M. Cruddas, and D.A. Compston, *Symptomatic Epstein-Barr virus infection and multiple sclerosis*. J Neurol Neurosurg Psychiatry, 1993. **56**(2): p. 167-8. PubMed ID: 8382268.
74. Haahr, S., et al., *Increased risk of multiple sclerosis after late Epstein-Barr virus infection: a historical prospective study*. Mult Scler, 1995. **1**(2): p. 73-7. PubMed ID: 9345455.
75. Marrie, R.A., et al., *Multiple sclerosis and antecedent infections: a case-control study*. Neurology, 2000. **54**(12): p. 2307-10. PubMed ID: 10881258.
76. Hernan, M.A., et al., *Multiple sclerosis and age at infection with common viruses*. Epidemiology, 2001. **12**(3): p. 301-6. PubMed ID: 11337603.
77. Goldacre, M.J., et al., *Multiple sclerosis after infectious mononucleosis: record linkage study*. J Epidemiol Community Health, 2004. **58**(12): p. 1032-5. PubMed ID: 15547068.
78. Thacker, E.L., F. Mirzaei, and A. Ascherio, *Infectious mononucleosis and risk for multiple sclerosis: a meta-analysis*. Ann Neurol., 2006. **59**(3): p. 499-503. PubMed ID: 16502434.
79. Nielsen, T.R., et al., *Multiple sclerosis after infectious mononucleosis*. Arch Neurol., 2007. **64**(1): p. 72-5. PubMed ID: 17210811.
80. Zaadstra, B.M., et al., *Selective association of multiple sclerosis with infectious mononucleosis*. Mult Scler., 2008. **14**(3): p. 307-13. Epub 2008 Jan 21. PubMed ID: 18208871.
81. Munch, M., et al., *The significance of Epstein-Barr virus seropositivity in multiple sclerosis patients?* Acta Neurol Scand, 1998. **97**(3): p. 171-4. PubMed ID: 9531433.
82. Pohl, D., et al., *High seroprevalence of Epstein-Barr virus in children with multiple sclerosis*. Neurology., 2006. **67**(11): p. 2063-5. PubMed ID: 17159123.
83. Lindsey, J.W., S. Patel, and J. Zou, *Epstein-Barr virus genotypes in multiple sclerosis*. Acta Neurol Scand., 2008. **117**(2): p. 141-4. PubMed ID: 18184350.
84. Bech, E., et al., *A randomized, double-blind, placebo-controlled MRI study of anti-herpes virus therapy in MS*. Neurology, 2002. **58**(1): p. 31-6. PubMed ID: 11781402.
85. Friedman, J.E., et al., *A randomized clinical trial of valacyclovir in multiple sclerosis*. Mult Scler., 2005. **11**(3): p. 286-95. PubMed ID: 15957509.
86. van Sechel, A.C., et al., *EBV-induced expression and HLA-DR-restricted presentation by human B cells of alpha B-crystallin, a candidate autoantigen in multiple sclerosis*. J Immunol, 1999. **162**(1): p. 129-35. PubMed ID: 9886378.
87. Rand, K.H., et al., *Molecular approach to find target(s) for oligoclonal bands in multiple sclerosis*. J Neurol Neurosurg Psychiatry, 1998. **65**(1): p. 48-55. PubMed ID: 9667560.
88. Esposito, M., et al., *Human transaldolase and cross-reactive viral epitopes identified by autoantibodies of multiple sclerosis patients*. J Immunol, 1999. **163**(7): p. 4027-32. PubMed ID: 10491006.

89. Vaughan, J.H., et al., *An Epstein Barr virus-related cross reactive autoimmune response in multiple sclerosis in Norway*. J Neuroimmunol, 1996. **69**(1-2): p. 95-102. PubMed ID: 8823380.
90. Lang, H.L., et al., *A functional and structural basis for TCR cross-reactivity in multiple sclerosis*. Nat Immunol, 2002. **3**(10): p. 940-3. Epub 2002 Sep 3. PubMed ID: 12244309.
91. Haahr, S., et al., *A putative new retrovirus associated with multiple sclerosis and the possible involvement of Epstein-Barr virus in this disease*. Ann N Y Acad Sci, 1994. **724**: p. 148-56. PubMed ID: 7518205.
92. Munch, M., et al., *B-lymphoblastoid cell lines from multiple sclerosis patients and a healthy control producing a putative new human retrovirus and Epstein-Barr virus*. Mult Scler, 1995. **1**(2): p. 78-81. PubMed ID: 9345456.
93. Christensen, T., et al., *Characterization of retroviruses from patients with multiple sclerosis*. Acta Neurol Scand Suppl, 1997. **169**: p. 49-58. PubMed ID: 9174640.
94. Christensen, T., et al., *Reverse transcriptase activity and particle production in B lymphoblastoid cell lines established from lymphocytes of patients with multiple sclerosis*. AIDS Res Hum Retroviruses, 1999. **15**(3): p. 285-91. PubMed ID: 10052759.
95. Pender, M.P., *Infection of autoreactive B lymphocytes with EBV, causing chronic autoimmune diseases*. Trends in Immunology, 2003. **24**(11): p. 584-8 PubMed ID: 14596882.
96. Warren, K.G., et al., *Isolation of Herpes simplex virus from human trigeminal ganglia, including ganglia from one patient with multiple sclerosis*. Lancet, 1977. **2**(8039): p. 637-9. PubMed ID: 71451.
97. Martin, J.R., R.K. Holt, and H.D. Webster, *Herpes-simplex-related antigen in human demyelinating disease and encephalitis*. Acta Neuropathol (Berl), 1988. **76**(4): p. 325-37. PubMed ID: 3176900.
98. Nicoll, J.A., E. Kinrade, and S. Love, *PCR-mediated search for herpes simplex virus DNA in sections of brain from patients with multiple sclerosis and other neurological disorders*. J Neurol Sci, 1992. **113**(2): p. 144-51. PubMed ID: 1336795.
99. Sanders, V.J., et al., *Herpes simplex virus in postmortem multiple sclerosis brain tissue*. Arch Neurol, 1996. **53**(2): p. 125-33. PubMed ID: 8639061.
100. Coyle, P.K. and P.A. Sibony, *Viral specificity of multiple sclerosis tear immunoglobulins*. J Neuroimmunol, 1987. **14**(2): p. 197-203. PubMed ID: 3029176.
101. Chou, Y.K., et al., *Frequency of T cells specific for myelin basic protein and myelin proteolipid protein in blood and cerebrospinal fluid in multiple sclerosis*. J Neuroimmunol, 1992. **38**(1-2): p. 105-13. PubMed ID: 1374422.
102. de Silva, S.M. and H.F. McFarland, *Multiple sclerosis patients have reduced HLA class II-restricted cytotoxic responses specific for both measles and herpes virus*. J Neuroimmunol, 1991. **35**(1-3): p. 219-26. PubMed ID: 1659588.
103. Brudek, T., et al., *Synergistic immune responses induced by endogenous retrovirus and herpesvirus antigens result in increased production of inflammatory cytokines in multiple sclerosis patients*. Scand J Immunol., 2008. **67**(3): p. 295-303. PubMed ID: 18261041.
104. Coyle, P.K. and Z. Procyk-Dougherty, *Multiple sclerosis immune complexes: an analysis of component antigens and antibodies*. Ann Neurol, 1984. **16**(6): p. 660-7. PubMed ID: 6098217.
105. Coyle, P.K., *CSF immune complexes in multiple sclerosis*. Neurology, 1985. **35**(3): p. 429-32. PubMed ID: 2579355.

106. Lycke, J., et al., *Acyclovir treatment of relapsing-remitting multiple sclerosis. A randomized, placebo-controlled, double-blind study.* J Neurol, 1996. **243**(3): p. 214-24. PubMed ID: 8936350.
107. Brudek, T., et al., *Simultaneous presence of endogenous retrovirus and herpes virus antigens has profound effect on cell-mediated immune responses: implications for multiple sclerosis.* AIDS Res Hum Retroviruses, 2004. **20**(4): p. 415-23. PubMed ID: 15157360.
108. Brudek, T., et al., *Activation of endogenous retrovirus reverse transcriptase in multiple sclerosis patient lymphocytes by inactivated HSV-1, HHV-6 and VZV.* J Neuroimmunol., 2007. **187**(1-2): p. 147-55. Epub 2007 May 10. PubMed ID: 17493688.
109. Perron, H., et al., *Herpes simplex virus ICP0 and ICP4 immediate early proteins strongly enhance expression of a retrovirus harboured by a leptomeningeal cell line from a patient with multiple sclerosis.* J Gen Virol, 1993. **74**(Pt 1): p. 65-72. PubMed ID: 7678635.
110. Cortese, I., et al., *Cross-reactive phage-displayed mimotopes lead to the discovery of mimicry between HSV-1 and a brain-specific protein.* J Neuroimmunol, 2001. **113**(1): p. 119-28. PubMed ID: 11137583.
111. Merelli, E., et al., *Human herpes virus 6 and human herpes virus 8 DNA sequences in brains of multiple sclerosis patients, normal adults and children.* J Neurol, 1997. **244**(7): p. 450-4. PubMed ID: 9266465.
112. Friedman, J.E., et al., *The association of the human herpesvirus-6 and MS.* Mult Scler, 1999. **5**(5): p. 355-62. PubMed ID: 10516780.
113. Goodman, A.D., et al., *Human herpesvirus 6 genome and antigen in acute multiple sclerosis lesions.* J Infect Dis, 2003. **187**(9): p. 1365-76. Epub 2003 Apr 15. PubMed ID: 12717617.
114. Opsahl, M.L. and P.G. Kennedy, *Early and late HHV-6 gene transcripts in multiple sclerosis lesions and normal appearing white matter.* Brain, 2005. **128**(Pt 3): p. 516-27. Epub 2005 Jan 19. PubMed ID: 15659422.
115. Tuke, P.W., et al., *Distribution and quantification of human herpesvirus 6 in multiple sclerosis and control brains.* Mult Scler, 2004. **10**(4): p. 355-9. PubMed ID: 15327029.
116. Mameli, G., et al., *Brains and peripheral blood mononuclear cells of multiple sclerosis (MS) patients hyperexpress MS-associated retrovirus/HERV-W endogenous retrovirus, but not Human herpesvirus 6.* J Gen Virol., 2007. **88**(Pt 1): p. 264-74. PubMed ID: 17170460.
117. Gutierrez, J., et al., *Multiple sclerosis and human herpesvirus 6.* Infection, 2002. **30**(3): p. 145-9. PubMed ID: 12120939.
118. Tejada-Simon, M.V., et al., *Detection of viral DNA and immune responses to the human herpesvirus 6 101-kilodalton virion protein in patients with multiple sclerosis and in controls.* J Virol, 2002. **76**(12): p. 6147-54. PubMed ID: 12021348.
119. Rotola, A., et al., *Human herpesvirus 6 infects the central nervous system of multiple sclerosis patients in the early stages of the disease.* Mult Scler, 2004. **10**(4): p. 348-54. PubMed ID: 15327028.
120. Alvarez-Lafuente, R., et al., *Human herpesvirus-6 and multiple sclerosis: relapsing-remitting versus secondary progressive.* Mult Scler., 2007. **13**(5): p. 578-83. Epub 2007 Feb 16. PubMed ID: 17548435.
121. Yao, K., et al., *Reactivation of human herpesvirus-6 in natalizumab treated multiple sclerosis patients.* PLoS ONE., 2008. **3**(4): p. e2028. PubMed ID: 18446218.

122. Ongradi, J., et al., *A pilot study on the antibodies to HHV-6 variants and HHV-7 in CSF of MS patients*. J Neurovirol, 1999. **5**(5): p. 529-32. PubMed ID: 10568890.
123. Soldan, S.S., et al., *Increased lymphoproliferative response to human herpesvirus type 6A variant in multiple sclerosis patients*. Ann Neurol, 2000. **47**(3): p. 306-13. PubMed ID: 10716249.
124. Pohl, D., et al., *Balo's concentric sclerosis associated with primary human herpesvirus 6 infection*. J Neurol Neurosurg Psychiatry., 2005. **76**(12): p. 1723-5. PubMed ID: 16291903.
125. Caselli, E., et al., *Detection of antibodies directed against human herpesvirus 6 U94/REP in sera of patients affected by multiple sclerosis*. J Clin Microbiol, 2002. **40**(11): p. 4131-7. PubMed ID: 12409386.
126. Virtanen, J.O., et al., *Evidence for human herpesvirus 6 variant A antibodies in multiple sclerosis: diagnostic and therapeutic implications*. J Neurovirol., 2007. **13**(4): p. 347-52. PubMed ID: 17849318.
127. Enbom, M., et al., *Similar humoral and cellular immunological reactivities to human herpesvirus 6 in patients with multiple sclerosis and controls*. Clin Diagn Lab Immunol, 1999. **6**(4): p. 545-9. PubMed ID: 10391860.
128. Tejada-Simon, M.V., et al., *Cross-reactivity with myelin basic protein and human herpesvirus-6 in multiple sclerosis*. Ann Neurol, 2003. **53**(2): p. 189-97. PubMed ID: 12557285.
129. Cirone, M., et al., *Human herpesvirus 6 and multiple sclerosis: a study of T cell cross-reactivity to viral and myelin basic protein antigens*. J Med Virol, 2002. **68**(2): p. 268-72. PubMed ID: 12210418.
130. Hong, J., et al., *Anti-viral properties of interferon beta treatment in patients with multiple sclerosis*. Mult Scler, 2002. **8**(3): p. 237-42. PubMed ID: 12120696.
131. Alvarez-Lafuente, R., et al., *Beta-interferon treatment reduces human herpesvirus-6 viral load in multiple sclerosis relapses but not in remission*. Eur Neurol, 2004. **52**(2): p. 87-91. Epub 2004 Jul 22. PubMed ID: 15273429.
132. Garcia-Montojo, M., et al., *Interferon beta treatment: bioavailability and antiviral activity in multiple sclerosis patients*. J Neurovirol., 2007. **13**(6): p. 504-12. PubMed ID: 18097882.
133. Kong, H., et al., *Human herpesvirus type 6 indirectly enhances oligodendrocyte cell death*. J Neurovirol, 2003. **9**(5): p. 539-50. PubMed ID: 13129768.
134. Soldan, S.S., et al., *Elevated serum and cerebrospinal fluid levels of soluble human herpesvirus type 6 cellular receptor, membrane cofactor protein, in patients with multiple sclerosis*. Ann Neurol, 2001. **50**(4): p. 486-93. PubMed ID: 11603380.
135. Chapenko, S., et al., *Correlation between HHV-6 reactivation and multiple sclerosis disease activity*. J Med Virol, 2003. **69**(1): p. 111-7. PubMed ID: 12436486.
136. Berti, R., et al., *Increased detection of serum HHV-6 DNA sequences during multiple sclerosis (MS) exacerbations and correlation with parameters of MS disease progression*. J Neurovirol, 2002. **8**(3): p. 250-6. PubMed ID: 12053279.
137. Alvarez-Lafuente, R., et al., *Relapsing-remitting multiple sclerosis and human herpesvirus 6 active infection*. Arch Neurol, 2004. **61**(10): p. 1523-7. PubMed ID: 15477505.
138. Alvarez-Lafuente, R., et al., *Clinical parameters and HHV-6 active replication in relapsing-remitting multiple sclerosis patients*. J Clin Virol., 2006. **37 Suppl 1**: p. S24-6. PubMed ID: 17276363.

139. Dietrich, J., et al., *Infection with an endemic human herpesvirus disrupts critical glial precursor cell properties*. J Neurosci., 2004. **24**(20): p. 4875-83. PubMed ID: 15152048.
140. Fotheringham, J., et al., *Human herpesvirus 6 (HHV-6) induces dysregulation of glutamate uptake and transporter expression in astrocytes*. J Neuroimmune Pharmacol., 2008. **3**(2): p. 105-16. Epub 2007 Sep 8. PubMed ID: 18247129.
141. Tait, A.R. and S.K. Straus, *Phosphorylation of U24 from Human Herpes Virus type 6 (HHV-6) and its potential role in mimicking myelin basic protein (MBP) in multiple sclerosis*. FEBS Lett., 2008. **582**(18): p. 2685-8. Epub 2008 Jul 9. PubMed ID: 18616943.
142. Hauser, S.L., et al., *Analysis of human T-lymphotropic virus sequences in multiple sclerosis tissue*. Nature, 1986. **322**(6075): p. 176-7. PubMed ID: 3014351.
143. Rozenberg, F., et al., *Analysis of retroviral sequences in the spinal form of multiple sclerosis*. Ann Neurol, 1991. **29**(3): p. 333-6. PubMed ID: 2042949.
144. Perron, H., et al., *Antibody to reverse transcriptase of human retroviruses in multiple sclerosis*. Acta Neurol Scand, 1991. **84**(6): p. 507-13. PubMed ID: 1724334.
145. Reddy, E.P., et al., *Amplification and molecular cloning of HTLV-I sequences from DNA of multiple sclerosis patients*. Science, 1989. **243**(4890): p. 529-33. PubMed ID: 2536193.
146. Greenberg, S.J., et al., *Detection of sequences homologous to human retroviral DNA in multiple sclerosis by gene amplification*. Proc Natl Acad Sci U S A, 1989. **86**(8): p. 2878-82. PubMed ID: 2468159.
147. Watanabe, T., et al., *No evidence of HTLV-1 infection in Japanese multiple sclerosis patients in polymerase chain reaction*. Jpn J Cancer Res, 1989. **80**(11): p. 1017-20. PubMed ID: 2514161.
148. Oksenberg, J.R., et al., *HTLV-I sequences are not detected in peripheral blood genomic DNA or in brain cDNA of multiple sclerosis patients*. Ann Neurol, 1990. **28**(4): p. 574-7. PubMed ID: 2252368.
149. Ehrlich, G.D., et al., *Multiple sclerosis, retroviruses, and PCR. The HTLV-MS Working Group*. Neurology, 1991. **41**(3): p. 335-43. PubMed ID: 1848687.
150. French, D., et al., *Amplifications of multiple regions of the HTLV-I genome from DNA of an Italian spastic paraparesis patient but not from DNA of multiple sclerosis patients*. J Neurol Sci, 1991. **103**(1): p. 82-9. PubMed ID: 1865236.
151. Merelli, E., et al., *Failure to detect genomic material of HTLV-I or HTLV-II in mononuclear cells of Italian patients with multiple sclerosis and chronic progressive myelopathy*. Eur Neurol, 1993. **33**(1): p. 23-6. PubMed ID: 8440281.
152. Kuroda, Y., et al., *Incidence of antibody to HTLV-I is not increased in Japanese MS patients*. Neurology, 1987. **37**(1): p. 156-8. PubMed ID: 3025773.
153. Brookes, S.M., et al., *The immune response to and expression of cross-reactive retroviral gag sequences in autoimmune disease*. Br J Rheumatol, 1992. **31**(11): p. 735-42. PubMed ID: 1280512.
154. Odum, N., et al., *HLA-DP antigens and HTLV-1 antibody status among Japanese with multiple sclerosis: evidence for an increased frequency of HLA-DPw4*. J Immunogenet, 1989. **16**(6): p. 467-73. PubMed ID: 2641759.
155. Ferrante, P., et al., *HTLV-I antibodies in multiple sclerosis and other neurological diseases*. Acta Neurol (Napoli), 1990. **12**(1): p. 95-9. PubMed ID: 2337003.
156. Banki, K., et al., *Human T-cell lymphotropic virus (HTLV)-related endogenous sequence, HRES-1, encodes a 28-kDa protein: a possible autoantigen for HTLV-*

- I gag-reactive autoantibodies*. Proc Natl Acad Sci U S A, 1992. **89**(5): p. 1939-43. PubMed ID: 1347429.
157. Birnbaum, G., S. Aubitz, and L. Kotilinek, *Search for autonomously proliferating spinal fluid lymphocytes in patients with multiple sclerosis*. Neurology, 1988. **38**(1): p. 28-30. PubMed ID: 3336460.
 158. Svenningsson, A., et al., *No evidence for spumavirus or oncovirus infection in relapsing-remitting multiple sclerosis*. Ann Neurol, 1992. **32**(5): p. 711-4. PubMed ID: 1333176.
 159. Nishimura, M., et al., *Human T lymphotropic virus type I (HTLV-I)-specific T helper cell responses from HTLV-I seronegative patients with chronic myelopathy and MS in Japan*. Mult Scler, 1996. **1**(4): p. 200-3. PubMed ID: 9345434.
 160. Sandberg-Wollheim, M., et al., *Bone marrow derived cells express human T-cell lymphotropic virus type I (HTLV-I)-related antigens in patients with multiple sclerosis*. Scand J Immunol, 1988. **28**(6): p. 801-6. PubMed ID: 3068791.
 161. Rasmussen, H.B., et al., *Expression of endogenous retroviruses in blood mononuclear cells and brain tissue from multiple sclerosis patients*. Mult Scler, 1995. **1**(2): p. 82-7. PubMed ID: 9345457.
 162. Rasmussen, H.B., et al., *Possible association between multiple sclerosis and the human T cell leukemia virus (HTLV)-related endogenous element, HRES-1*. Mult Scler, 1996. **2**(3): p. 133-6. PubMed ID: 9345377.
 163. Rasmussen, H.B., et al., *Haplotypes of the endogenous retrovirus HRES-1 in multiple sclerosis patients and healthy control subjects of Shanghai Chinese origin*. Dis Markers, 1998. **13**(4): p. 251-5. PubMed ID: 9553740.
 164. Rasmussen, H.B. and J. Clausen, *A novel haplotype of the endogenous retrovirus, HRES-1, in patients with multiple sclerosis and healthy individuals*. Autoimmunity, 1999. **29**(2): p. 141-5. PubMed ID: 10433076.
 165. Rasmussen, H.B., et al., *Association between the endogenous retrovirus HRES-1 and multiple sclerosis in the United Kingdom--evidence of genetically different disease subsets?* Dis Markers, 2000. **16**(3-4): p. 101-4. PubMed ID: 11381188.
 166. Rasmussen, H.B., et al., *Three allelic forms of the human endogenous retrovirus, ERV3, and their frequencies in multiple sclerosis patients and healthy individuals*. Autoimmunity, 1996. **23**(2): p. 111-7. PubMed ID: 8871766.
 167. Rasmussen, H.B. and J. Clausen, *Large number of polymorphic nucleotides and a termination codon in the env gene of the endogenous human retrovirus ERV3*. Dis Markers, 1998. **14**(3): p. 127-33. PubMed ID: 10427470.
 168. Christensen, T., et al., *Expression of sequence variants of endogenous retrovirus RGH in particle form in multiple sclerosis*. Lancet, 1998. **352**(9133): p. 1033. PubMed ID: 9759750.
 169. Christensen, T., et al., *Molecular characterization of HERV-H variants associated with multiple sclerosis*. Acta Neurol Scand, 2000. **101**(4): p. 229-38. PubMed ID: 10770518.
 170. Christensen, T., et al., *Antibodies against a human endogenous retrovirus and the preponderance of env splice variants in multiple sclerosis patients*. Mult Scler, 2003. **9**(1): p. 6-15. PubMed ID: 12617261.
 171. Antony, J.M., et al., *Human endogenous retrovirus glycoprotein-mediated induction of redox reactants causes oligodendrocyte death and demyelination*. Nat Neurosci, 2004. **7**(10): p. 1088-95. Epub 2004 Sep 26. PubMed ID: 15452578.
 172. Johnston, J.B., et al., *Monocyte activation and differentiation augment human endogenous retrovirus expression: implications for inflammatory brain diseases*. Ann Neurol, 2001. **50**(4): p. 434-42. PubMed ID: 11601494.

173. Muradrasoli, S., et al., *Development of real-time PCRs for detection and quantitation of human MMTV-like (HML) sequences HML expression in human tissues.* J Virol Methods., 2006. **136**(1-2): p. 83-92. Epub 2006 May 19. PubMed ID: 16713632.
174. Moyes, D.L., et al., *HERV-K113 is not associated with multiple sclerosis in a large family-based study.* AIDS Res Hum Retroviruses., 2008. **24**(3): p. 363-5. PubMed ID: 18327982.
175. Moyes, D.L., et al., *The distribution of the endogenous retroviruses HERV-K113 and HERV-K115 in health and disease.* Genomics., 2005. **86**(3): p. 337-41. PubMed ID: 16024218.
176. Tai, A.K., et al., *Human endogenous retrovirus-K18 Env as a risk factor in multiple sclerosis.* Mult Scler, 2008. **13**: p. 13 PubMed ID: 18701576.
177. Perron, H., et al., *Molecular identification of a novel retrovirus repeatedly isolated from patients with multiple sclerosis. The Collaborative Research Group on Multiple Sclerosis.* Proc Natl Acad Sci U S A., 1997. **94**(14): p. 7583-8. PubMed ID: 9207135.
178. Menard, A., et al., *Gliotoxicity, reverse transcriptase activity and retroviral RNA in monocyte/macrophage culture supernatants from patients with multiple sclerosis.* FEBS Lett, 1997. **413**(3): p. 477-85. PubMed ID: 9303559.
179. Serra, C., et al., *Multiple sclerosis and multiple sclerosis-associated retrovirus in Sardinia.* Neurol Sci, 2001. **22**(2): p. 171-3. PubMed ID: 11603622.
180. Nowak, J., et al., *Multiple sclerosis-associated virus-related pol sequences found both in multiple sclerosis and healthy donors are more frequently expressed in multiple sclerosis patients.* J Neurovirol, 2003. **9**(1): p. 112-7. PubMed ID: 12587074.
181. de Villiers, J.N., et al., *Analysis of viral and genetic factors in South African patients with multiple sclerosis.* Metab Brain Dis., 2006. **21**(2-3): p. 163-9. Epub 2006 Jul 22. PubMed ID: 16865539.
182. Dolei, A., et al., *Multiple sclerosis-associated retrovirus (MSRV) in Sardinian MS patients.* Neurology, 2002. **58**(3): p. 471-3. PubMed ID: 11839854.
183. Petzold, A., et al., *No evidence for MSRV viraemia and glial cell death in acute optic neuritis.* Mult Scler., 2005. **11**(4): p. 495-6. PubMed ID: 16042236.
184. Antony, J.M., et al., *Quantitative analysis of human endogenous retrovirus-W env in neuroinflammatory diseases.* AIDS Res Hum Retroviruses., 2006. **22**(12): p. 1253-9. PubMed ID: 17209768.
185. Antony, J.M., et al., *Comparative expression of human endogenous retrovirus-W genes in multiple sclerosis.* AIDS Res Hum Retroviruses., 2007. **23**(10): p. 1251-6. PubMed ID: 17961112.
186. Mameli, G., et al., *Inhibition of multiple-sclerosis-associated retrovirus as biomarker of interferon therapy.* J Neurovirol., 2008. **14**(1): p. 73-7. PubMed ID: 18300077.
187. Sotgiu, S., et al., *Multiple sclerosis-associated retrovirus in early multiple sclerosis: a six-year follow-up of a Sardinian cohort.* Mult Scler., 2006. **12**(6): p. 698-703. PubMed ID: 17262996.
188. Sotgiu, S., et al., *Multiple sclerosis-associated retrovirus and optic neuritis.* Mult Scler., 2006. **12**(3): p. 357-9. PubMed ID: 16764351.
189. Perron, H., et al., *Human endogenous retrovirus (HERV)-W ENV and GAG proteins: physiological expression in human brain and pathophysiological modulation in multiple sclerosis lesions.* J Neurovirol., 2005. **11**(1): p. 23-33. PubMed ID: 15804956.

190. Zawada, M., et al., *MSRV pol sequence copy number as a potential marker of multiple sclerosis*. Pol J Pharmacol, 2003. **55**(5): p. 869-75. PubMed ID: 14704480.
191. Christensen, T., et al., *Gene-environment interactions in multiple sclerosis: innate and adaptive immune responses to human endogenous retrovirus and herpesvirus antigens and the lectin complement activation pathway*. J Neuroimmunol., 2007. **183**(1-2): p. 175-88. Epub 2006 Nov 16. PubMed ID: 17113160.
192. Rolland, A., et al., *Correlation between disease severity and in vitro cytokine production mediated by MSRV (multiple sclerosis associated retroviral element) envelope protein in patients with multiple sclerosis*. J Neuroimmunol, 2005. **160**(1-2): p. 195-203. Epub 2004 Dec 8. PubMed ID: 15710473.
193. Ruprecht, K., et al., *Lack of immune responses against multiple sclerosis-associated retrovirus/human endogenous retrovirus W in patients with multiple sclerosis*. J Neurovirol., 2008. **14**(2): p. 143-51. PubMed ID: 18444086.
194. Perron, H., et al., *Multiple sclerosis retrovirus particles and recombinant envelope trigger an abnormal immune response in vitro, by inducing polyclonal Vbeta16 T-lymphocyte activation*. Virology, 2001. **287**(2): p. 321-32. PubMed ID: 11531410.
195. Firouzi, R., et al., *Multiple sclerosis-associated retrovirus particles cause T lymphocyte-dependent death with brain hemorrhage in humanized SCID mice model*. J Neurovirol, 2003. **9**(1): p. 79-93. PubMed ID: 12587071.
196. Chesters, P.M., J. Heritage, and D.J. McCance, *Persistence of DNA sequences of BK virus and JC virus in normal human tissues and in diseased tissues*. J Infect Dis, 1983. **147**(4): p. 676-84. PubMed ID: 6302172.
197. Stoner, G.L., et al., *Immunocytochemical search for JC papovavirus large T-antigen in multiple sclerosis brain tissue*. Acta Neuropathol, 1986. **70**(3-4): p. 345-7. PubMed ID: 3532688.
198. Buckle, G.J., et al., *Lack of JC viral genomic sequences in multiple sclerosis brain tissue by polymerase chain reaction*. Ann Neurol, 1992. **32**(6): p. 829-31. PubMed ID: 1335225.
199. Ferrante, P., et al., *Detection of JC virus DNA in cerebrospinal fluid from multiple sclerosis patients*. Mult Scler, 1998. **4**(2): p. 49-54. PubMed ID: 9599333.
200. Alvarez-Lafuente, R., et al., *JC virus in cerebrospinal fluid samples of multiple sclerosis patients at the first demyelinating event*. Mult Scler., 2007. **13**(5): p. 590-5. Epub 2007 Feb 16. PubMed ID: 17548437.
201. Agostini, H.T., et al., *Influence of JC virus coding region genotype on risk of multiple sclerosis and progressive multifocal leukoencephalopathy*. J Neurovirol, 2000. **6 Suppl 2**: p. S101-8. PubMed ID: 10871796.
202. Bogdanovic, G., et al., *Detection of JC virus in cerebrospinal fluid (CSF) samples from patients with progressive multifocal leukoencephalopathy but not in CSF samples from patients with herpes simplex encephalitis, enteroviral meningitis, or multiple sclerosis*. J Clin Microbiol, 1998. **36**(4): p. 1137-8. PubMed ID: 9542955.
203. Yousry, T.A., et al., *Evaluation of patients treated with natalizumab for progressive multifocal leukoencephalopathy*. N Engl J Med., 2006. **354**(9): p. 924-33. PubMed ID: 16510746.
204. Franciotta, D., et al., *Failure to detect JC virus DNA in cerebrospinal fluid of multiple sclerosis patients*. Mult Scler., 2006. **12**(5): p. 674-5. PubMed ID: 17086918.
205. Delbue, S., et al., *JC virus viremia in interferon-beta -treated and untreated Italian multiple sclerosis patients and healthy controls*. J Neurovirol, 2007. **13**(1): p. 73-7. PubMed ID: 17454451.

206. Alvarez-Lafuente, R., et al., *Interferon-beta treatment and active replication of the JC virus in relapsing-remitting multiple sclerosis patients*. Eur J Neurol., 2007. **14**(2): p. 233-6. PubMed ID: 17250736.
207. Sindic, C.J., et al., *Detection of CSF-specific oligoclonal antibodies to recombinant JC virus VP1 in patients with progressive multifocal leukoencephalopathy*. J Neuroimmunol, 1997. **76**(1-2): p. 100-4. PubMed ID: 9184638.
208. Du Pasquier, R.A., et al., *JC virus induces a vigorous CD8+ cytotoxic T cell response in multiple sclerosis patients*. J Neuroimmunol., 2006. **176**(1-2): p. 181-6. Epub 2006 Jun 5. PubMed ID: 16750575.
209. Rastogi, S.C., et al., *Partial purification of MS specific brain antigens*. Acta Neurol Scand, 1979. **59**(6): p. 281-96. PubMed ID: 90442.
210. Dowling, P.C., et al., *Measles virus nucleic acid sequences in human brain*. Virus Res, 1986. **5**(1): p. 97-107. PubMed ID: 3751288.
211. Woyciechowska, J.L., D.L. Madden, and J.L. Sever, *Absence of measles-virus antigen in jejunum of multiple-sclerosis patients*. Lancet, 1977. **2**(8047): p. 1046-9. PubMed ID: 72954.
212. Gupta, J.K., et al., *Multiple sclerosis and malabsorption*. Am J Gastroenterol, 1977. **68**(6): p. 560-5. PubMed ID: 612212.
213. Brankin, B., et al., *Failure to detect measles virus RNA, by reverse transcription-polymerase chain reaction, in peripheral blood leucocytes of patients with multiple sclerosis*. Mult Scler, 1996. **1**(4): p. 204-6. PubMed ID: 9345435.
214. Fewster, M.E., F.R. Ames, and M.C. Botha, *Measles antibodies and histocompatibility types in multiple sclerosis*. J Neurol Sci, 1979. **43**(1): p. 19-26. PubMed ID: 521827.
215. Rostrom, B., et al., *Viral antibody activity of oligoclonal and polyclonal immunoglobulins synthesized within the central nervous system in multiple sclerosis*. Ann Neurol, 1981. **9**(6): p. 569-74. PubMed ID: 7259119.
216. Schadlich, H.J., et al., *Intrathecal synthesis of virus antibodies: a diagnostic test for multiple sclerosis*. Eur Neurol, 1990. **30**(5): p. 302-4. PubMed ID: 2269323.
217. Reiber, H., S. Ungefehr, and C. Jacobi, *The intrathecal, polyspecific and oligoclonal immune response in multiple sclerosis*. Mult Scler, 1998. **4**(3): p. 111-7. PubMed ID: 9762657.
218. Rostrom, B., *Antibodies against viruses and structural brain components in oligoclonal IgG obtained from multiple sclerosis brain*. J Neurol, 1982. **226**(4): p. 255-63. PubMed ID: 6174706.
219. Bollengier, F., et al., *Multiple sclerosis: oligoclonal IgG, kappa-lambda light chain distribution and measles antibodies in brain extracts*. Brain Res, 1978. **152**(1): p. 133-44. PubMed ID: 98212.
220. Neighbour, P.A., A.E. Miller, and B.R. Bloom, *Interferon responses of leukocytes in multiple sclerosis*. Neurology, 1981. **31**(5): p. 561-6. PubMed ID: 6164954.
221. Jacobson, S., M.L. Flerlage, and H.F. McFarland, *Impaired measles virus-specific cytotoxic T cell responses in multiple sclerosis*. J Exp Med, 1985. **162**(3): p. 839-50. PubMed ID: 2411841.
222. Santoli, D., et al., *Cytotoxic activity and interferon production by lymphocytes from patients with multiple sclerosis*. J Immunol, 1981. **126**(4): p. 1274-8. PubMed ID: 6162885.
223. Walker, J.E., et al., *HLA and the response of lymphocytes to viral antigens in patients with multiple sclerosis*. Hum Immunol, 1982. **4**(1): p. 71-8. PubMed ID: 7061242.

224. Salonen, R., et al., *Defective production of interferon-alpha associated with HLA-DW2 antigen in stable multiple sclerosis*. J Neurol Sci, 1982. **55**(2): p. 197-206. PubMed ID: 6982311.
225. Makela, M.J., *Antibody response to different antigenic sites on measles virus surface polypeptides in patients with multiple sclerosis*. J Neurol Sci, 1989. **90**(2): p. 239-46. PubMed ID: 2470864.
226. Ovsyannikova, I.G., et al., *HLA class II alleles and measles virus-specific cytokine immune response following two doses of measles vaccine*. Immunogenetics, 2005. **56**(11): p. 798-807. Epub 2005 Jan 27. PubMed ID: 15712014.
227. Haile, R., et al., *A study of measles virus and canine distemper virus antibodies, and of childhood infections in multiple sclerosis patients and controls*. J Neurol Sci, 1982. **56**(1): p. 1-10. PubMed ID: 7143023.
228. Sullivan, C.B., B.R. Visscher, and R. Detels, *Multiple sclerosis and age at exposure to childhood diseases and animals: cases and their friends*. Neurology, 1984. **34**(9): p. 1144-8. PubMed ID: 6540400.
229. Bachmann, S. and J. Kesselring, *Multiple sclerosis and infectious childhood diseases*. Neuroepidemiology, 1998. **17**(3): p. 154-60. PubMed ID: 9648121.
230. Casetta, I., et al., *Environmental risk factors and multiple sclerosis: a community-based, case-control study in the province of Ferrara, Italy*. Neuroepidemiology, 1994. **13**(3): p. 120-8. PubMed ID: 8015665.
231. Bager, P., et al., *Childhood infections and risk of multiple sclerosis*. Brain, 2004. **127**(Pt 11): p. 2491-7. Epub 2004 Sep 15. PubMed ID: 15371288.
232. Svenningsson, A., et al., *Incidence of MS during two fifteen-year periods in the Gothenburg region of Sweden*. Acta Neurol Scand, 1990. **82**(3): p. 161-8. PubMed ID: 2270743.
233. Matossian-Rogers, A., A. Dos Santos, and H. Festenstein, *Human cytotoxic T-cells against measles virus-infected and myelin basic protein-coated targets are cross-reactive*. Int Arch Allergy Appl Immunol, 1987. **84**(2): p. 159-64. PubMed ID: 2443455.
234. Richert, J.R., et al., *Measles virus-specific human T cell clones: studies of alloreactivity and antigenic cross-reactivity*. J Neuroimmunol, 1988. **19**(1-2): p. 59-68. PubMed ID: 2456307.
235. Pette, M., et al., *Measles virus-directed responses of CD4+ T lymphocytes in MS patients and healthy individuals*. Neurology, 1993. **43**(10): p. 2019-25. PubMed ID: 8105423.
236. Jingwu, Z., et al., *Antibodies to myelin basic protein and measles virus in multiple sclerosis: precursor frequency analysis of the antibody producing B cells*. Autoimmunity, 1991. **11**(1): p. 27-34. PubMed ID: 1725965.
237. Rubio, N. and A. Cuesta, *Lack of cross-reaction between myelin basic proteins and putative demyelinating virus envelope proteins*. Mol Immunol, 1989. **26**(7): p. 663-8. PubMed ID: 2476671.
238. Godec, M.S., et al., *Absence of measles, mumps, and rubella viral genomic sequences from multiple sclerosis brain tissue by polymerase chain reaction*. Ann Neurol, 1992. **32**(3): p. 401-4. PubMed ID: 1416811.
239. Berr, C., et al., *Risk factors in multiple sclerosis: a population-based case-control study in Hautes-Pyrenees, France*. Acta Neurol Scand, 1989. **80**(1): p. 46-50. PubMed ID: 2782041.
240. Sindic, C.J., P. Monteyne, and E.C. Laterre, *The intrathecal synthesis of virus-specific oligoclonal IgG in multiple sclerosis*. J Neuroimmunol, 1994. **54**(1-2): p. 75-80. PubMed ID: 7523446.

241. Ilonen, J., et al., *Lymphocyte blast transformation responses and viral antibodies in relation to HLA antigens in multiple sclerosis*. J Neurol Sci, 1981. **49**(1): p. 117-33. PubMed ID: 6259296.
242. Hays, P., *Multiple sclerosis and delayed mumps*. Acta Neurol Scand, 1992. **85**(3): p. 200-3. PubMed ID: 1575004.
243. Pekmezovic, T., M. Jarebinski, and J. Drulovic, *Childhood infections as risk factors for multiple sclerosis: Belgrade case-control study*. Neuroepidemiology, 2004. **23**(6): p. 285-8. PubMed ID: 15297795.
244. Bansil, S., et al., *Multiple sclerosis in India: a case-control study of environmental exposures*. Acta Neurol Scand, 1997. **95**(2): p. 90-5. PubMed ID: 9059727.
245. Tanaka, R., Y. Iwasaki, and H. Koprowski, *Paramyxovirus-like structures in brains of multiple sclerosis patients*. Arch Neurol, 1975. **32**(2): p. 80-3. PubMed ID: 1122181.
246. Hayano, M., J.H. Sung, and A.R. Mastri, *"Paramyxovirus-like" intranuclear inclusions occurring in the nervous system in diverse unrelated conditions*. J Neuropathol Exp Neurol, 1976. **35**(3): p. 287-94. PubMed ID: 178836.
247. Kirk, J. and W.M. Hutchinson, *The fine structure of the CNS in multiple sclerosis. I. Interpretation of cytoplasmic papovavirus-like and paramyxovirus-like inclusions*. Neuropathol Appl Neurobiol, 1978. **4**(5): p. 343-56. PubMed ID: 724090.
248. ter Meulen, V., et al., *Fusion of cultured multiple-sclerosis brain cells with indicator cells: presence of nucleocapsids and virions and isolation of parainfluenza-type virus*. Lancet, 1972. **2**(7766): p. 1-5. PubMed ID: 4113621.
249. Cernescu, C., F. Verdes, and Y. Sorodoc, *The incidence of antiviral antibodies in multiple sclerosis*. Virologie, 1977. **28**(4): p. 251-8. PubMed ID: 203087.
250. Whitaker, J.N., et al., *Immunogenetic analysis and serum viral antibody titers in multiple sclerosis*. Arch Neurol, 1976. **33**(6): p. 399-403. PubMed ID: 938263.
251. Goswami, K.K., et al., *Antibodies against the paramyxovirus SV5 in the cerebrospinal fluids of some multiple sclerosis patients*. Nature, 1987. **327**(6119): p. 244-7. PubMed ID: 3553964.
252. Vandvik, B. and E. Norrby, *Paramyxovirus SV5 and multiple sclerosis*. Nature, 1989. **338**(6218): p. 769-71. PubMed ID: 2541340.
253. McLean, B.N. and E.J. Thompson, *Antibodies against the paramyxovirus SV5 are not specific for cerebrospinal fluid from multiple sclerosis patients*. J Neurol Sci, 1989. **92**(2-3): p. 261-6. PubMed ID: 2809621.
254. Ciongoli, A.K., et al., *In vitro cellular responsiveness in multiple sclerosis patients to a viral isolate from multiple sclerosis brain tissue and to other antigens*. Neurology, 1975. **25**(9): p. 891-3. PubMed ID: 169493.
255. Utermohlen, V. and J.B. Zabriskie, *A suppression of cellular immunity in patients with multiple sclerosis*. J Exp Med, 1973. **138**(6): p. 1591-6. PubMed ID: 4357685.
256. Rauch, H.C., K.M. King, and L.J. Lewandowski, *Detection of cellular hypersensitivity among multiple sclerosis (MS) patients to 6/94 virus; a parainfluenza type 1 isolate from MS brain tissue*. Int Arch Allergy Appl Immunol, 1975. **48**(4): p. 475-84. PubMed ID: 164416.
257. Vervliet, G., et al., *Interferon production and natural killer (NK) activity in leukocyte cultures from multiple sclerosis patients*. J Neurol Sci, 1983. **60**(1): p. 137-50. PubMed ID: 6192218.
258. Stoger, I., et al., *Lack of correlation between impaired interferon production and natural killer activity of lymphocytes in multiple sclerosis*. Arch Virol, 1982. **71**(3): p. 259-65. PubMed ID: 6179500.

259. Vervliet, G., et al., *Interferon production by cultured peripheral leucocytes of MS patients*. Clin Exp Immunol, 1984. **58**(1): p. 116-26. PubMed ID: 6206970.
260. Rostasy, K., et al., *Chlamydia pneumoniae in children with MS: frequency and quantity of intrathecal antibodies*. Neurology, 2003. **61**(1): p. 125-8. PubMed ID: 12847174.
261. Ciongoli, A.K., et al., *In vitro cellular responsiveness in multiple sclerosis patients to a purified measles virus nuclear core and to other viral antigens*. J Neurol Sci, 1976. **28**(3): p. 331-8. PubMed ID: 932780.
262. Compston, D.A., et al., *Viral infection in patients with multiple sclerosis and HLA-DR matched controls*. Brain, 1986. **109**(Pt 2): p. 325-44. PubMed ID: 3456817.
263. Ito, M., et al., *Antibody titers by mixed agglutination to varicella-zoster, herpes simplex and vaccinia viruses in patients with multiple sclerosis*. Proc Soc Exp Biol Med, 1975. **149**(4): p. 835-9. PubMed ID: 170625.
264. Cremer, N.E., et al., *Comprehensive viral immunology of multiple sclerosis. II. Analysis of serum and CSF antibodies by standard serologic methods*. Arch Neurol, 1980. **37**(10): p. 610-5. PubMed ID: 6252874.
265. Weiner, H.L., J. Cherry, and K. McIntosh, *Decreased lymphocyte transformation to vaccinia virus in multiple sclerosis*. Neurology, 1978. **28**(5): p. 415-20. PubMed ID: 565482.
266. Fuccillo, D.A., et al., *Multiple sclerosis: cellular and humoral immune responses to several viruses*. Neurology, 1978. **28**(6): p. 613-5. PubMed ID: 206860.
267. Greenstein, J.I., et al., *The lymphoproliferative response to measles virus in twins with multiple sclerosis*. Ann Neurol, 1984. **15**(1): p. 79-87. PubMed ID: 6608918.
268. Ordonez, G., et al., *Brief presence of varicella-zoster viral DNA in mononuclear cells during relapses of multiple sclerosis*. Arch Neurol, 2004. **61**(4): p. 529-32. PubMed ID: 15096401.
269. Sotelo, J., et al., *Varicella-zoster virus in cerebrospinal fluid at relapses of multiple sclerosis*. Ann Neurol., 2008. **63**(3): p. 303-11. PubMed ID: 18306233.
270. Myhr, K.M., et al., *Altered antibody pattern to Epstein-Barr virus but not to other herpesviruses in multiple sclerosis: a population based case-control study from western Norway*. J Neurol Neurosurg Psychiatry, 1998. **64**(4): p. 539-42. PubMed ID: 9576551.
271. Forghani, B., et al., *Comprehensive viral immunology of multiple sclerosis. III. Analysis of CSF antibodies by radioimmunoassay*. Arch Neurol, 1980. **37**(10): p. 616-9. PubMed ID: 6252875.
272. Bednarova, J., P. Stourac, and P. Adam, *Relevance of immunological variables in neuroborreliosis and multiple sclerosis*. Acta Neurol Scand., 2005. **112**(2): p. 97-102. PubMed ID: 16008535.
273. Ross, R.T. and M. Cheang, *Geographic similarities between varicella and multiple sclerosis: an hypothesis on the environmental factor of multiple sclerosis*. J Clin Epidemiol, 1995. **48**(6): p. 731-7. PubMed ID: 7769403.
274. Perez-Cesari, C., M.M. Saniger, and J. Sotelo, *Frequent association of multiple sclerosis with varicella and zoster*. Acta Neurol Scand., 2005. **112**(6): p. 417-9. PubMed ID: 16281927.
275. Ross, R.T., L.E. Nicolle, and M. Cheang, *The varicella zoster virus: a pilot trial of a potential therapeutic agent in multiple sclerosis*. J Clin Epidemiol, 1997. **50**(1): p. 63-8. PubMed ID: 9048691.
276. Schmutzhard, E., P. Pohl, and G. Stanek, *Borrelia burgdorferi antibodies in patients with relapsing/remitting form and chronic progressive form of multiple sclerosis*. J Neurol Neurosurg Psychiatry, 1988. **51**(9): p. 1215-8. PubMed ID: 3225603.

277. Chmielewska-Badora, J., E. Cisak, and J. Dutkiewicz, *Lyme borreliosis and multiple sclerosis: any connection? A seroepidemic study*. Ann Agric Environ Med, 2000. **7**(2): p. 141-3. PubMed ID: 11153045.
278. Weigelt, W., T. Schneider, and R. Lange, *Sequence homology between spirochaete flagellin and human myelin basic protein*. Immunol Today, 1992. **13**(7): p. 279-80. PubMed ID: 1382434.
279. Karussis, D., H.L. Weiner, and O. Abramsky, *Multiple sclerosis vs Lyme disease: a case presentation to a discussant and a review of the literature*. Mult Scler, 1999. **5**(6): p. 395-402. PubMed ID: 10618695.
280. Morre, S.A., et al., *Is Chlamydia pneumoniae present in the central nervous system of multiple sclerosis patients?* Ann Neurol, 2000. **48**(3): p. 399. PubMed ID: 10976651.
281. Ke, Z., et al., *Lack of detectable Chlamydia pneumoniae in brain lesions of patients with multiple sclerosis*. Ann Neurol, 2000. **48**(3): p. 400. PubMed ID: 10976653.
282. Hammerschlag, M.R., et al., *Is Chlamydia pneumoniae present in brain lesions of patients with multiple sclerosis?* J Clin Microbiol, 2000. **38**(11): p. 4274-6. PubMed ID: 11060110.
283. Sriram, S., et al., *Detection of chlamydial bodies and antigens in the central nervous system of patients with multiple sclerosis*. J Infect Dis., 2005. **192**(7): p. 1219-28. Epub 2005 Sep 2. PubMed ID: 16136465.
284. Saiz, A., et al., *No evidence of CNS infection with Chlamydia pneumoniae in patients with multiple sclerosis*. J Neurol, 2001. **248**(7): p. 617-8. PubMed ID: 11518005.
285. Derfuss, T., et al., *Intrathecal antibody production against Chlamydia pneumoniae in multiple sclerosis is part of a polyspecific immune response*. Brain, 2001. **124**(Pt 7): p. 1325-35. PubMed ID: 11408328.
286. Budak, F., et al., *The investigation of Chlamydia pneumoniae in patients with multiple sclerosis*. Int J Neurosci., 2007. **117**(3): p. 409-15. PubMed ID: 17365124.
287. Lindsey, J. and S. Patel, *PCR for bacterial 16S ribosomal DNA in multiple sclerosis cerebrospinal fluid*. Mult Scler., 2008. **14**(2): p. 147-52. Epub 2007 Nov 6. PubMed ID: 17986505.
288. Sotgiu, S., et al., *Chlamydia pneumoniae in the cerebrospinal fluid of patients with multiple sclerosis and neurological controls*. Mult Scler, 2001. **7**(6): p. 371-4. PubMed ID: 11795458.
289. Hao, Q., et al., *Chlamydia pneumoniae infection associated with enhanced MRI spinal lesions in multiple sclerosis*. Mult Scler, 2002. **8**(5): p. 436-40. PubMed ID: 12356213.
290. Contini, C., et al., *Cerebrospinal fluid molecular demonstration of Chlamydia pneumoniae DNA is associated to clinical and brain magnetic resonance imaging activity in a subset of patients with relapsing-remitting multiple sclerosis*. Mult Scler, 2004. **10**(4): p. 360-9. PubMed ID: 15327030.
291. Yamamoto, Y., *PCR in diagnosis of infection: detection of bacteria in cerebrospinal fluids*. Clin Diagn Lab Immunol, 2002. **9**(3): p. 508-14. PubMed ID: 11986253.
292. Sessa, R., et al., *Real time PCR for detection of Chlamydia pneumoniae in peripheral blood mononuclear cells of patients with multiple sclerosis*. J Neurol, 2007. **26**: p. 26 PubMed ID: 17460814.

293. Krametter, D., et al., *Chlamydia pneumoniae in multiple sclerosis: humoral immune responses in serum and cerebrospinal fluid and correlation with disease activity marker*. Mult Scler, 2001. **7**(1): p. 13-8. PubMed ID: 11321187.
294. Parratt, J., et al., *Chlamydia pneumoniae-specific serum immune complexes in patients with multiple sclerosis*. Mult Scler., 2008. **14**(3): p. 292-9. Epub 2008 Jan 21. PubMed ID: 18208884.
295. Sriram, S., et al., *Chlamydia pneumoniae infection of the central nervous system in multiple sclerosis*. Ann Neurol, 1999. **46**(1): p. 6-14. PubMed ID: 10401775.
296. Krametter, D., et al., *Chlamydia pneumoniae-specific humoral immune responses and clinical disease parameters in multiple sclerosis*. Ann Neurol, 2001. **49**(1): p. 135. PubMed ID: 11198285.
297. Munger, K.L., et al., *A prospective study of Chlamydia pneumoniae infection and risk of MS in two US cohorts*. Neurology, 2004. **62**(10): p. 1799-803. PubMed ID: 15159481.
298. Yao, S.Y., et al., *CSF oligoclonal bands in MS include antibodies against Chlamydophila antigens*. Neurology, 2001. **56**(9): p. 1168-76. PubMed ID: 11342681.
299. Sriram, S., W. Mitchell, and C. Stratton, *Multiple sclerosis associated with Chlamydia pneumoniae infection of the CNS*. Neurology, 1998. **50**(2): p. 571-2. PubMed ID: 9484408.
300. Sriram, S., et al., *Pilot study to examine the effect of antibiotic therapy on MRI outcomes in RRMS*. J Neurol Sci., 2005. **234**(1-2): p. 87-91. PubMed ID: 15935383.
301. Woessner, R., et al., *Long-term antibiotic treatment with roxithromycin in patients with multiple sclerosis*. Infection., 2006. **34**(6): p. 342-4. PubMed ID: 17180590.
302. Alonso, A., et al., *Antibiotic use and risk of multiple sclerosis*. Am J Epidemiol., 2006. **163**(11): p. 997-1002. Epub 2006 Apr 5. PubMed ID: 16597708.
303. MacIntyre, A., et al., *Chlamydia pneumoniae infection alters the junctional complex proteins of human brain microvascular endothelial cells*. FEMS Microbiol Lett, 2002. **217**(2): p. 167-72. PubMed ID: 12480099.
304. Lenz, D.C., et al., *A Chlamydia pneumoniae-specific peptide induces experimental autoimmune encephalomyelitis in rats*. J Immunol, 2001. **167**(3): p. 1803-8. PubMed ID: 11466406.
305. Grimaldi, L.M., et al., *An MRI study of Chlamydia pneumoniae infection in Italian multiple sclerosis patients*. Mult Scler, 2003. **9**(5): p. 467-71. PubMed ID: 14582771.
306. Contini, C., et al., *Chlamydophila pneumoniae DNA and mRNA transcript levels in peripheral blood mononuclear cells and cerebrospinal fluid of patients with multiple sclerosis*. Neurosci Res., 2008. **62**(1): p. 58-61. Epub 2008 May 20. PubMed ID: 18572268.
307. Buljevac, D., et al., *Chlamydia pneumoniae and the risk for exacerbation in multiple sclerosis patients*. Ann Neurol, 2003. **54**(6): p. 828-31. PubMed ID: 14681894.